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(54) Title: CHEMOKINE-LIKE PROTEINS AND ME	THODS	OF USE
(57) Abstract		
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"CHEMOKINE-LIKE PROTEINS AND METHODS OF USE"

Background of the Invention

The invention relates to novel chemokine-like

proteins and their use to suppress the proliferation of
actively dividing myeloid cells, e.g., myeloid progenitor
cells, myeloid stem cells, and leukemic cells, and to
mobilize progenitor cells from the bone marrow to the
peripheral blood.

Each year approximately 173,000 of the people who undergo chemotherapy become neutropenic, which causes 10 them to become susceptible to infection and anemia (Hecht, Drug and Market Development, 4:49, 1993). One method of treatment for neutropenia, e.g., chemotherapy-15 induced neutropenia, includes stimulation of progenitor cells with differentiation factors including granulocyte macrophage-colony stimulating factor (GM-CSF), granulocyte-colony stimulating factor (G-CSF), and erythropoietin (EPO) as a method of salvaging cells 20 surviving chemotherapy. Approximately 100,000 patients annually are suitable for receiving G-CSF for this purpose. G-CSF is used to stimulate growth of white blood cell progenitors, and EPO has been used to stimulate production of red blood cells, however, there 25 are no megakaryocyte progenitor stimulating factors

currently available.

An alternative approach for preventing neutropenia is to inhibit cell proliferation with low doses of various chemokines, which inhibits cell cycling, thereby protecting the progenitor cells from the effects of chemotherapy and/or radiation therapy. After chemotherapy has ended, the chemokine treatment is also stopped, which allows the progenitor cells to resume normal proliferation.

Chemokines are small inducible proteins that are related by amino acid homology, chromosome location, and structural similarities, including the presence of four position-invariant cysteine residues in their primary amino acid sequence that form two disulfide bonds. The amino acid sequences of various naturally occurring, wild-type chemokine proteins are shown in Fig. 1.

Certain chemokines, known as beta chemokines, have a Cys-Cys pair as the first two cysteines, and include

10 macrophage inflammatory protein-1 alpha (MTP-1a), MIP-1\(\beta\), macrophage chemotactic and activating factor (MCAF, also known as monocyte chemo-attractant protein-1 (MCP-1), MCP-3, and Regulated on Activation, Normal T-cell Expressed and Secreted protein (RANTES). The beta

15 chemokines are potent chemoattractants for a variety of blood cell components, including monocytes, eosinophils, and T-lymphocytes, but not neutrophils.

other chemokines, known as alpha chemokines, have a Cys-X-Cys triplet as the first two cysteines (X can be 20 any amino acid other than cysteine), and include the human-derived proteins interleukin-8 (IL-8), GRO-α (also called melanoma-growth stimulating activity (MGSA/GRO), MIP-2β (also known as GRO-β), MIP-2β (also known as GRO-γ), neutrophil activating peptide-2 (NAP-2), platelet factor 4 (PF4), gamma interferon inducible protein 10 (γIP-10), Epithelial derived Neutrophil activating protein (78 amino acids in length) (ENA-78), β-thromboglobulin (βTG), connective tissue-activating peptide-III (CTAP-III), and platelet basic protein (PBP).

The alpha chemokines are potent chemoattractants and all except PF4 and γIP-10 activate neutrophils.

Chemokines have been shown to regulate proliferation and/or differentiation of hematopoietic stem and progenitor cells in <u>vitro</u> and in <u>vivo</u>. For 35 example, MIP-1a, YIP-10, IL-8, Gro-\$, PF4, and MCP-1 have

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been shown to inhibit the proliferation of colony forming unit-granulocyte macrophage (CFU-GM), burst forming unit-erythroid cells (BRU-E), and colony forming unit-multipotential progenitor cells (CFU-GEMM), at concentrations greater than 25 ng/ml when administered to mice. See, e.g., Broxmeyer et al., J. Immunol., 150:3448-3458 (1993).

On the other hand, several members of the chemokine family, including NAP-2, Gro-α, Gro-γ, RANTES, and MIP-1β have been shown not to possess such inhibitory activities, but Gro-α and Gro-γ have been shown to interfere with the inhibitory activity of IL-8 and PF4. Furthermore, Broxmeyer et al., WO 94/13321, states that combinations of any two of MCAF, MIP-1α, MIP-2α, IL-8, γIP-10 and PF4 provide a decrease in the concentrations required for suppressing progenitor cell proliferation.

Several groups have also examined the ability of chemokines to inhibit the proliferation of progenitor cells in vivo. For example, Maze et al., J. Immunol., 20 149:1004 (1992), observed that murine MIP-10 suppressed proliferation and absolute numbers of granulocytemacrophage, erythroid, and multipotential progenitor cells from mice femurs and spleens at doses between 2 and 10 µg per mouse, injected intravenously. Murine MIP-16, which was unable to inhibit cell proliferation in vitre, displayed no biological activity in vivo either.

Recombinant human chemokines have also been demonstrated to inhibit proliferation of CFU-GM, CFU-GEMM, and BFU-E following intravenous injection into mice. Furthermore, active chemokines cause a significant decrease in the number of progenitor cells in S-phase. For example, Dunlop et al., Blood, 79:2221 (1992), observed that recombinant human MIP-10 was able to suppress CFU-S in a dose dependent manner in vitro, and

to reduce the high proliferative state of the CFU-S compartment to a quiescent state in vivo.

Human MIP-1α has been shown to protect progenitor cells in vivo from the cytotoxic effects of the chemotherapeutic drug cytosine arabinoside (ARA-C), and Caen et al., Blood, 82:162a (1993), has reported that PF4, at doses between 1 and 5 μg/mouse, protected hematopoietic precursor cells from the adverse effects of the chemotherapeutic, 5-fluorouracil. Lord et al., Blood, 79:2605-2609 (1992), also observed that MIP-1α protected myeloid progenitors in a murine system from the cytotoxic effects of hydroxyurea.

In addition, Gewirtz et al., <u>J. Clin. Invest.</u>, <u>68</u>:56 (1989), observed that a peptide from PF4 containing the C-terminal 24 amino acids inhibited proliferation of megakaryocyte progenitors in <u>vitro</u> at a concentration of 25 μg/ml. A shorter peptide containing only the last 13 amino acids from PF4 was found to be inactive in the assay. Similarly, a peptide containing the C-terminal 18 amino acids from β-thromboglobulin did not inhibit proliferation. Recently, Caen et al., <u>Blood</u>, <u>82</u>:162a (1933), reported that a dodecapeptide, Asn-Gly-Arg-Lys-

proliferation. Receively, the provided Ash-Gly-Arg-Lys-(1993), reported that a dodecapeptide, Ash-Gly-Arg-Lys-Ile-Cys-Leu-Asp-Leu-Glu-Ala-Pro, which is able to inhibit human and murine megakaryocyte and platelet production, human and protect hematopoietic precursor cells in vivo (1-5 µg/mouse) during 5-fluorouracil chemotherapy.

Although G-CSFs are the drugs most commonly used to treat chemically-induced neutropenia, they have certain drawbacks. For example, G-CSFs do not prevent a drop in white blood cell count, i.e., they do not avoid neutropenia, but merely shorten the low point or nadir in the blood count. G-CSFs also fail to stimulate platelet development, and thus do not protect platelets.

Moreover, G-CSFs are expensive, and are therefore often

administered only after the white cell count drops below about 1000.

Summary of the Invention

The present invention is based on the discovery 5 that the ability of chemokines to suppress the proliferation of actively dividing myeloid cells (e.g., myeloid progenitor cells, myeloid stem cells, and leukemic cells) is associated with the presence of ϵ : least two "active domains." "Active domains" are defined 10 herein as regions of several contiguous amino acids in chemokines that, in combinations of two or more active domains, are necessary for myelosuppressive activity. The present inventors have determined the amino acid sequences of five of these active domains: 1) ELR 15 (derived from IL-8), 2) ELRV (derived from IL-8), 3) DLQ (one of the two DLQ domains from PF4), 4) IATLKNGQK (SEQ ID NO:43) (derived from $Gro\beta$), and 5) ACLNPASPIVK (SEQ ID NO:44) (derived from Groα). Various mutations of these sequences can serve as these active domains.

Based on these discoveries, the invention includes novel chemokine-like proteins that are based upon a wild-20 type chemokine amino acid sequence (the "template" chemokine) that is modified to include at least one heterologous active domain, e.g., an active domain from a 25 chemokine other than the template chemokine. The novel chemokine-like proteins thus contain at least two active domains, and exhibit myelosuppressive activity (i.e., suppress the proliferation of actively dividing myeloid cells). Certain of these new chemokine-like proteins

30 have a much higher myelosuppressive activity than naturally occurring, wild-type chemokines.

Further based on these discoveries, the invention features the new proteins and the use of those proteins to treat chemotherapy-induced, as well as radiationinduced, neutropenia. The invention also features new methods of screening and treating patients with certain myelogenous leukemias as well as other hyperproliferative blood diseases. In addition, the new proteins can be used to identify, purify, and expand progenitor-specific cell populations in an ex vivo setting for reintroduction into a patient following radiation or chemotherapy.

In general, the invention features a protein 10 having the amino acid sequence of a wild-type chemokine having four cysteine residues, the amino acid sequence containing an active domain selected from: ELR, DLN, ELQ, DLR, ELRV, DLQ, IATLKNGQK (SEQ ID NO:43), and ACLNPASPIVK (SEQ ID NO:44). Preferably, the active 15 domain replaces a number of amino acids identical to the number of amino acids in the active domain, and is positioned in the protein such that the active domain is: 1) on the N-terminus side of and proximal the first cysteine residue of the wild-type chemokine; 2) on the N-20 terminus side of and within 11 amino acids of the third cysteine residue of the wild-type chemokine; 3) on the Nterminus side of and within 11 amino acids of the fourth cysteine residue of the wild-type chemokine; or 4) on the C-terminus side of and within 11 amino acids of the 25 fourth cysteine residue of the wild-type chemokine.

The invention also features a protein having the amino acid sequence of a wild-type chemokine, e.g., IL-8, having four cysteine residues, with the following modifications: a) amino acids DLQ as the 3 contiguous amino acids on the N-terminus side of and proximal to the first cysteine residue of the wild-type chemokine; and b) amino acids ELXVX₁X₂X₃X₄X₅X₆ as the 10 amino acids on the N-terminus side of and proximal to the third cysteine of the wild-type chemokine, wherein X is any amino acid, 35 e.g., R, N, E, or Q, and X₁X₂X₃X₄X₅X₆ can be any six

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consecutive amino acids on the N-terminus side of and proximal to the third cysteine of the wild-type chemokine. In particular, the protein can have the amino acid sequence SAKDLQCQCIKTYSKPF-

5 HPKFIKELRVIESGPHCANTEIIVKLSDGRELCLDPKENWVQRVVEKFLKRAENS (SEQ ID NO:16).

The invention additionally features a protein having the amino acid sequence of a wild-type chemokine, e.g., IL-8, having four cysteine residues, with the 10 following modifications: a) amino acids ELQ as the 3 contiguous amino acids on the N-terminus side of and proximal to the first cysteine residue of the wild-type chemokine; and b) amino acids $\text{ELXVX}_1 X_2 X_3 X_4 X_5 X_6$ as the 10 amino acids on the N-terminus side of and proximal to the 15 third cysteine of the wild-type chemokine, wherein X is any amino acid, e.g., R, N, E, or Q, and $X_1X_2X_3X_4X_5X_6$ can be any six consecutive amino acids on the N-terminus side of and proximal to the third cysteine of the wild-type chemokine. In particular, the protein can have the amino 20 acid sequence

SAKELQCQCIKTYSKPFHPKFIKELRVIESGPHCANTEIIVKLSDGRELCLDPKENW VQRVVEKFLKRAENS (SEQ ID NO:47).

The invention additionally features a protein having the amino acid sequence of a wild-type chemokine, 25 e.g., IL-8, having four cysteine residues, with the following modifications: a) amino acids DLR as the 3 contiguous amino acids on the N-terminus side of and proximal to the first cysteine residue of the wild-type chemokine; and b) amino acids $ELXVX_1X_2X_3X_4X_5X_6$ as the 10 30 amino acids on the N-terminus side of and proximal to the third cysteine of the wild-type chemokine, wherein X is any amino acid, e.g., R, N, E, or Q, and $X_1X_2X_3X_4X_5X_6$ can be any six consecutive amino acids on the N-terminus side of and proximal to the third cysteine of the wild-type 35 chemokine. In particular, the protein can have the amino

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(54) Title: CHEMOKINE-LIKE PROTEINS AND METHODS OF USE

(57) Abstract

The invention relates to novel chemokine-like proteins that include two or more newly discovered active domains from different chemokines. Active domains are regions of several contiguous amino acids that are necessary for chemokines' ability to suppress the chemokines actively dividing myeloid cells, e.g., myeloid progenitor cells, myeloid stem cells, and leuternic cells. The new chemokine-like proteins provide higher myelosuppressive activity than naturally occurring, wild-type chemokines.

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following modifications: a) amino acids DLQ as the 3 contiguous amino acids on the N-terminus side of and proximal to the first cysteine residue of the wild-type chemokine; and b) amino acids ACLNPASPIVK (SEQ ID NO:44) 5 replacing 11 amino acids including the fourth cysteine of the wild-type chemokine, wherein the C in ACLNPASPIVK corresponds to the fourth cysteine residue. In particular, the protein can have the amino acid sequence EAEEDGDLQCLCVKTTSQVRPRHITSLEVIKAGPH-

10 CPTAQLIATLKNGRKACLNPASPIVKKIIKKLLES (SEQ ID NO:26).

The invention further features a protein having the amino acid sequence of a wild-type chemokine, e.g. PF4, having four cysteine residues, with the following modifications: a) amino acids DLQ as the 3 contiguous 15 amino acids on the N-terminus side of and proximal to the first cysteine residue of the wild-type chemokine; and b) amino acids IATLKNGQK (SEQ ID NO:43) and Z as the 10 amino acids on the N-terminus side of and proximal to the fourth cysteine of the wild-type chemokine, wherein Z is 20 any amino acid, e.g., I, A, L, or R. In particular, the protein can have the amino acid sequence

EAEEDGDLQCLCVKTTSQVQPQHITSLEV-

IKAGPHCPTAQLIATLKNGQKICLDLQAPLYKKIIKKLLES (SEQ ID NO:28).

In any of these new proteins, the wild-type 25 chemokine can be, e.g., IL-8, GRO- α , MIP- 2α , MIP- 1α , PF4,

NAP-2, ENA-78, PBP, CTAP-III, β TG, γ IP-10, MCAF, or PANTES.

The invention also feature a protein having the amino acid sequence SAKELRCQCIKTYSKPFHPKFIKE-

30 YRRIESGPHCANTEIIVKLSDGRELCLDPKENWVQRVVEKFL (SEQ ID NO:17).

The invention also features the use of these proteins to suppress proliferation of an actively dividing myeloid cell by contacting the cell with an 35 effective amount of any of the new chemokine-like

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proteins of the invention. The cell can be cultured in witro, or ex vivo, or the cell can be in vivo. The proteins can also be used for the manufacture of a medicament for treating a hyperproliferative myeloid disease or as an adjunctive agent, e.g., in chemotherapy.

Furthermore, the invention features an adjunctive method for use with chemotherapy or radiation therapy in a patient by administering an effective amount of a chemokine-like protein of the invention to the patient, and administering chemotherapy or radiation therapy to

the patient in conjunction with the administration of the protein. As used herein, "in conjunction with" means before, during, or after, or any combination thereof.

The invention also features these proteins for use in treating a hyperproliferative myeloid disease, e.g., chronic myelogenous leukemia, polycythemia vera, or a hypermegakaryocytopoietic disorder, in a patient by administering to the patient an effective amount of a chemokine-like protein of the invention.

20 In addition, the invention features methods of detecting and/or isolating CD34* myeloid cells in a sample of cells by obtaining a sample of cells, contacting the sample with a chemokine-like protein under conditions that allow the protein to bind to any CD34*

25 myeloid cells in the sample to form bound complexes, and detecting any bound complexes in the sample as an indication of the presence of CD34⁺ myeloid cells in the sample, or removing any bound complexes from the sample, and separating CD34⁺ myeloid cells from the bound
30 complexes to isolate the CD34⁺ myeloid cells from the

The invention additionally features a method of isolating CD34⁺ myeloid cells from the peripheral blood of a patient by administering an effective amount of a chemokine-like protein to the patient, and isolating

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CD34+ myeloid cells from a peripheral blood sample from the patient.

The invention further features nucleic acids encoding the chemokine-like proteins of the invention, 5 such as nucleic acids having the sequence of SEQ ID NO:34 or SEQ ID NO:41.

Unless otherwise defined, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art 10 to which this invention belongs. Although methods and materials similar or equivalent to those described herein can be used in the practice or testing of the present invention, the preferred methods and materials are described below. All publications, patent applications, patents, and other references mentioned herein are incorporated by reference. In addition, the materials, methods, and examples are illustrative only and not intended to be limiting.

Other features and advantages of the invention

20 will be apparent from the detailed description including
the drawings, and from the claims.

<u>Drawings</u>

Fig. 1 is a representation of the wild-type amino acid sequences of representative members of the human 25 chemokine family of proteins (SEQ ID NOS:1-15).

Fig. 2 is a representation of the amino acid sequences of novel chemokine-like proteins of the invention (SEQ ID NOS:16-28 and 47-49) compared to the wild-type amino acid sequences of IL-8 (SEQ ID NO:1) and 30 PF4 (SEQ ID NO:6).

Fig. 3A is a representation of the nucleotide sequence of human PF4 cDNA (SEQ ID NO:29) and the translated amino acid sequence (SEQ ID NOS:66 AND 6), as

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well as the sequences of the four amplimers used for its synthesis (SEQ ID NOS:30-33).

Fig. 3B is a representation of the nucleotide sequence of the cDNA of new chemokine-like protein PF4M2 5 (SEQ ID NO:34) and the translated amino acid sequence, as well as the sequences of the two amplimers used for its synthesis (SEQ ID NOS:33 and 35).

Fig. 3C is a representation of the nucleotide sequence (SEQ ID NO:26, which begins at nucleotide 10 residue number 101 of SEQ ID NO:57) of the cDNA of new chemokine-like protein PF4-414 (SEQ ID NO:57) and the translated amino acid sequence, as well as the sequences of the amplimers used for its synthesis (SEQ ID NOS:58 and 30-32).

15 Fig. 3D is a representation of the nucleotide sequence of the cDNA of new chemokine-like protein PF4-426 (SEQ ID NO:59) and the translated amino acid sequence, as well as the sequences of the amplimers used for its synthesis (SEQ ID NOS:33 and 60-62).

Fig. 4A is a representation of the nucleotide sequence of human IL-8 cDNA (SEQ ID NO:36) and the translated amino acid sequence (SEQ ID NO:69), as well as the sequences of the four amplimers used for its synthesis (SEQ ID NOS:37-40).

25 Fig. 4B is a representation of the nucleotide sequence of the cDNA of new chemokine-like protein IL-8M1 (SEQ ID NO:41) and the translated amino acid sequence (SEQ ID NO:70), as well as the sequences of the two amplimers used for its synthesis (SEQ ID NOS:40 and 42).

Fig. 4C is a representation of the nucleotide sequence of the cDNA of the IL-8 polypeptide antagonist IL-8M8 (SEQ ID NO:52) and its translated amino acid sequence (SEQ ID NO:63), as well as the two amplimers used for its synthesis (SEQ ID NOS:40 and 53).

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Fig. 4D is a representation of the nucleotide sequence of the cDNA of the IL-8 polypeptide antagonist IL-8M9 (SEQ ID NO:54) and its translated amino acid sequence (SEQ ID NO:64), as well as the two amplimers 5 used for its synthesis (SEQ ID NOS:40 and 55).

Fig. 4E is a representation of the nucleotide sequence of the cDNA of the IL-8 polypeptide antagonist IL-8M10 (SEQ ID NO:50) and its translated amino acid sequence (SEQ ID NO:65), as well as the two amplimers 10 used for its synthesis (SEQ ID NOS:40 and 51).

Fig. 4F is a representation of the nucleotide sequence encoding IL-8M3 (SEQ ID NO:56).

Fig. 5 is a schematic of the chemokine active domains necessary for myelosuppression and their locations in wild-type IL-8 and PF4, and in new chemokine-like proteins.

Figs. 6A to 6H are graphs representing the inhibition of myeloid progenitor colony formation in in witro assays of new chemokine-like proteins compared to wild-type PF4 and IL-8.

Fig. 7A is a bar graph comparing the ability of the new chemokine-like proteins to elicit neutrophil degranulation as detected by release of elastase activity from neutrophils.

25 Fig. 7B is a graph comparing elastase release activity of IL-8M1, IL-8M8, IL-8M9, IL-8M10, and wildtype IL-8 in an in vitro assay.

Fig. 8A is a bar graph comparing the influence of wild-type IL-8 and PF4 and new chemokine-like protein IL-30 8M1 on the cycling status of granulocyte-macrophage progenitor cells in vivo.

Fig. 8B is a bar graph comparing the influence of wild-type IL-8 and PF4 and new chemokine-like protein IL-8M1 on the absolute numbers of granulocyte-macrophage 35 progenitor cells in <u>vivo</u>.

Detailed Description

A series of new chemokine-like proteins were made using wild-type amino acid sequences of known chemokines as templates, and including at least two active domains, 5 with at least one of the new active domains originating from a chemokine other than the template chemokine. Other new proteins that included only one active domain were used as controls. These new chemokine-like proteins were then tested for myelosuppressive activity in in 10 vitro assays and in in vivo animal models described below. In addition, several of these new proteins were found not to activate neutrophils, and thus are not inflammatory.

The amino acid sequences of the new chemokine-like 15 proteins are shown in Fig. 2, in which the bold amino acids represent active domains that have been discovered to be required for the myelosuppressive activity of the wild-type chemokines and new chemokine-like proteins. Underlined amino acids are different from those normally 20 found at that location in the wild-type chemokine template. Variations of these new chemokine-like proteins can be made and tested as described below.

Chemokines

Chemokines having normal, wild-type amino acid 25 sequences are available in natural or recombinant form. For example, natural PF4 can be isolated from human platelet extracts. Natural IL-8 has been isolated from either stimulated human endothelial cells or human monocytes. Other chemokines such as PBP or β TG can be 30 isolated from various human cell sources. The wild-type amino acid sequences of several representative members of the human chemokine family are shown in Fig. 1. As used herein, the term "chemokine" or "wild-type chemokine"

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includes both the natural and recombinant forms of the proteins that have a wild-type amino acid sequence.

Recombinant MIP-1 α (rMIP-1 α), rMIP-1 β , rMIP-2 α , and rMIP-2 β are produced using yeast expression vectors

- and rmir-2p are produced and start start and rmir-2b as described in Tekamp-Olson et al., <u>J. Exp. Med.</u>, <u>172</u>:911 (1990), or can be obtained from Peprotech (Rocky Hills, NJ). Recombinant IL-8 (rIL-8), PF4 (rPF4), NAP-2 (rNAP-2), CTAP-III (rCTAP-III), GROα (rGRO-α), and GROβ (rGROβ) were cloned and expressed in <u>E. coli</u> at Repligen
- 10 (Cambridge, MA). Both the 77- and 72-amino acid forms of IL-8 were cloned and expressed at Repligen, and can be purchased from Pepro Tech, Rocky Hills, NJ. The 77-amino acid form of rIL-8 and the natural form of PF4 can also be purchased from Sigma Chemical Co. (St. Louis, MO).
- 15 Recombinant preparations of IL-8, MCAF and RANTES are available from Pepro Tech (Rocky Hills, NJ). Recombinant NAP-2 (rNAP-2) can be purchased from Bachem Bioscience, Philadelphia, PA. Other cytokines, including recombinant EPO (rEPO) can be purchased from Amgen Corporation
- 20 (Thousand Oaks, CA). Human rGM-CSF, rGROα, IL-3, and SLF (also called mast cell growth factor, stem cell factor, and c-kit ligand) can be obtained from Immunex Corporation (Seattle, WA).

Human chemokines were purified prior to use and no 25 endotoxin was detected in these samples using a standard limulus lysate assay.

Human Bone Marrow Progenitor Cells

Human bone marrow cells were obtained by aspiration from the posterior iliac crest of healthy volunteers. Low density (LD) cells (<1.077 g/cm³) were retrieved after density cut separation on FICOLL-HYPAQUE* (Pharmacia Fine Chemicals, Piscataway, NJ).

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Methods of Making Chemokine-Like Proteins

New chemokine-like proteins can be made using standard synthetic techniques, or can be generated using recombinant DNA technology and expressed in bacterial, 5 yeast, or mammalian cells using standard techniques.

Chemical Synthesis

Chemokine-like proteins can be synthesized based on the amino acid sequences described herein and variations thereof by standard solid-phase methods using 10 the tert-butyloxy-carbonyl and benzyl protection strategy described in Clark-Lewis et al., P.N.A.S., USA, 90:3574-3577 (1993) and Clark-Lewis et al., Biochemistry, 30:3128-3135 (1991). After deprotection with hydrogen fluoride, the proteins are folded by air oxidation and 15 purified by reverse-phase HPLC. Purity is determined by reverse-phase HPLC and isoelectric focusing. Amino acid incorporation is monitored during synthesis, and the final composition is determined by amino acid analysis. The correct covalent structure of the protein can be 20 confirmed using ion-spray mass spectrometry (SCIEX

APIII). Recombinant Expression of Chemokine-Like Proteins New chemokine-like proteins can also be generated using recombinant DNA techniques. For example, mutant 25 chemokine genes were generated using standard polymerase chain reaction (PCR) amplification of synthetic oligonucleotide primers, e.g., as described in Mullis et al., U.S. Patent No. 4,800,159. The primers were designed to be less than 100 bases in length with a 30 minimal overlap of at least 25 bases, and were synthesized using an Applied Biosystems 394 DNA/RNA synthesizer. A large (10 to 40 base) region upstream of a restriction site was included to facilitate restriction endonuclease digestion and to enhance visualization of 35 endonuclease digested DNA versus undigested DNA on an

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agarose gel. Oligonucleotide primers were amplified by 25 to 30 cycles of PCR using standard reaction conditions. Outside primer concentration was 0.4 pmoles/µl and all inside primer concentrations were 0.004 pmoles/µl in the reaction. Following amplification, the selected gene was digested with restriction endonucleases Bam HI and Nde I and electrophoresed on a 1% LMP agarose/3% Nusieve gel containing 40 mM Tris, acetate, 2 mM Na₂EDTA+2H₂O (TAE) buffer. Bands corresponding to the ligation reactions.

The digested DNA was ligated into the pMEK vector (derivatized pET 9a vector, Novagen, Madison, WI) and was selected for by kanamycin resistance. This vector permits protein expression only following induction of the cells with isopropyl-β-D-thiogalactoside (IPTG). Following the ligation reaction, the vector including the gene segment was electroporated into XL1 Blue E. Coli cells and screened on agar-kanamycin plates. Colonies were chosen and the correct inserts were screened by restriction digests of DNA mini-preps (5 colonies per plate). The DNA sequences of positive clones were confirmed using an Applied Biosystems Prism Taq Dye Deoxy Terminator sequencing kit and an Applied Biosystems 373λ

25 DNA sequencer.

Following sequence confirmation, the resulting plasmids were electroporated into BL21 (DE3) E. colicells. Colonies were screened on kanamycin plates and grown in 50 ml cultures for examination of expression levels following induction with IPTG.

Platelet Factor 4 (PF4)

The following general methods were used to synthesize, clone, and express a PF4 gene. First, the gene for PF4 was obtained by directing the bacterial 35 expression of the protein. As shown in Fig. 3A (SEQ ID

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NO:29, / = signal peptide cleavage site), (SEQ ID NO:66 is the amino acid sequence of the signal peptide) four partially complementary oligonucleotides (amplimers) were synthesized (PF4-1 to PF4-4). Amplimer PF4-1 (SEQ ID 5 NO:30) was a 99 mer and included an Nde I restriction site, the ATG translation start codon, and sense strand nucleotides 101 to 157 of the published human PF4 cDNA (Poncz et al., Blood, 69:219-223 (1987)) [codons Glu32 to Val_{50} of the precursor (Glu_1 to Val_{19} of the mature 10 secreted form)]. PF4-2 (SEQ ID NO:31), also a 99 mer, contained the antisense nucleotides 231 to 133, extending to the first two positions of codon $\ensuremath{\mathrm{Thr}}_{75}$ of the precursor $(\mathrm{Thr}_{44}\ \mathrm{of}\ \mathrm{the}\ \mathrm{secreted}\ \mathrm{form})$, with the 3' 25 nucleotides being complementary to the 3' 25 nucleotides of PF4-1. 15 PF4-3 (SEQ ID NO:32), also a 99 mer, contained sense nucleotides 205 to 303, extending to the coding strand sequence of Leu $_{99}$ of the precursor (Leu $_{68}$ of the mature secreted form), with positions 1 through 27 being complementary to positions 1 though 27 of PF4-2. PF4-4 20 (SEQ ID NO:33), a 70 mer, contained antisense nucleotides 313 to 274, extending to the ${ t TAG}_{102}$ translation stop of the precursor (TAG $_{71}$ of the mature secreted form), and contained additional nucleotides for restriction sites for cloning. The sequence of these amplimers were such 25 that the final double-stranded DNA fragment would encode the amino acid sequence of human PF4 as shown in Fig. 3A

(SEQ ID NO:29).

The four amplimers were annealed to create a gapped FF4 gene, and PCR amplification was then used to fill in the gaps and amplify the full-length gene so that it could be cloned into an expression vector for production of the protein. The four amplimers were mixed at final concentrations of 5 µM of PF4-1, 0.05 µM of PF4-2, 0.05 µM of PF4-3, and 5 µM of PF4-4. This mixture was diluted 6.25-fold and amplified using Pfu polymerase

(Stratagene, La Jolla, CA) per the manufacturer's conditions without adding any template in addition to the amplimer mixture. The reaction was subjected to 35 cycles of amplification at 98°C for 1 minute, 50°C for 1 minute, and 72°C for 3 minutes. The success of the amplification was verified by agarose gel electrophoresis of a portion of the PCR reaction mixture.

The amplified PF4 DNA fragment was purified and concentrated from the remainder of the reaction using a 10 Quiagen PCR Purification kit (Quiagen, Inc., Chatsworth, CA). One half of the purified PF4 DNA fragment was digested with 60 units each of the restriction enzymes NdeI and BamHI in a 50 μl reaction volume. The digested fragment was purified by low melting point agarose gel 15 electrophoresis and ligated with gel purified NdeI/BamHIdigested pET3a (Novagen, Madison, WI) to create the E. coli expression plasmid pETPF4-1. The pET3a backbone was derived from the plasmid pARVH which consisted of the PET3a vector with an MGSA/Gro-related sequence cloned in 20 the NdeI to BamHI restriction site. This plasmid was digested with NdeI and BamHI to remove the MGSA/Gro fragment so that the PF4 fragment could be inserted. The PF4 DNA fragment insert of the resulting pETPF4-1 plasmid was sequenced in its entirety, which confirmed that the 25 correct sequence was present.

For initial expression, the plasmid pETFF4-1 was used to transform <u>E. coli</u> strain BL21. The transformants were grown to an OD of 1.2, and then induced to express the PF4 gene by the addition of 0.4 mM IPTG to the culture medium. After 2 hours, the cells were pelleted, and a small portion was examined by SDS-PAGE and Western blotting to confirm expression of PF4.

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Interleukin-8

The gene for IL-8 was synthesized in a similar fashion except that the oligonucleotide amplimers were made such that the final sequence would encode the IL-8 5 amino acids (SEQ ID NO:69) as in Pig. 4A (SEQ ID NO:36). Amplimer IL-8-1 (SEQ ID NO:37) was a 99 mer and included an NdeI restriction site, the ATG translation start codon, and sense strand nucleotides 1-59 of the human IL-8 cDNA. IL-8-2 (SEQ ID NO:38) was a 99 mer and included 10 antisense nucleotides 42 to 140 of the human IL-8 cDNA. IL-8-3 (SEQ:ID NO:39), also a 99 mer, included sense strand nucleotides 109 to 207. IL-8-4 (SEQ ID NO:40), was a 70 mer, and included anti-sense strand nucleotides 183 to 219 of the IL-8 cDNA extending to the TAG 15 translation stop of the gene and an additional stretch of nucleotides for restriction sites for cloning.

Purification The synthetic genes for human PF4, IL-8, and new chemokine-like proteins were expressed in E. coli (BL21) 20 cells and grown in 10 liter containers. Cells were grown for 4 hours at 37°C followed by induction with IPTG (0.4 mM) overnight. Approximately 550 g of cell paste containing inclusion bodies was obtained, of which 100 g was suspended in lysis buffer (0.05 M Tris-HCl, pH 8.0, 5 25 mM EDTA, 5 mM DTT, 0.1% Triton-X100) and lysed by DYNOMIL* disruption in the presence of 0.1% α toluenesulfonylfluoride (PMSF). The lysed preparation was treated with two aliquots of DNase I, concomitantly added with MgCl2 and incubated for 0.5 hours at 4°C. Following DNase I treatment, the suspension was 30

centrifuged overnight at 4°C at 13,000 rpm. The precipitate from the centrifugation step was extracted in 150 mls of 0.05 M Tris-HCl, pH 8.0, 6 M Guanidine-HCl, and 50 mM DTT overnight at 25°C. The extracted material 35 was dialyzed against buffer containing 25 mM sodium

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acetate, pH 4.0, 8 M urea, and 50 mM DTT and loaded onto a S-Sepharose column equilibrated with 25 mM sodium acetate, pH 4.0, 8 M urea. The column, containing bound protein, was washed with 25 mM sodium acetate, pH 4.0, to 5 remove the urea followed by a second buffer wash containing 25 mM sodium acetate, pH 4.0, 0.5 M NaCl. The protein was then eluted with buffer containing 50 mM Tris-HCl, pH 8.0, 1 M NaCl.

Fractions containing the appropriate chemokine or 10 chemokine-like protein were refolded by overnight incubation in the presence of 1 mM oxidized/2 mM reduced glutathione at 25°C. The extent of refolding of the proteins was monitored using POROS* analytical chromatography. The reduced protein species were 15 observed to elute from the POROS™ column at different acetonitrile concentrations relative to the refolded species. Refolded fractions were pooled and rechromatographed by C4 semi-preparative reverse phase HPLC using a 0 to 100% acetonitrile gradient in 0.1% 20 TFA:H2O. Peak fractions were pooled and lyophilized for concentration determination.

Purity of the final proteins was assessed by Coomassie staining of SDS PAGE, analytical C4 reverse phase HPLC, and amino acid analysis. Protein 25 concentrations were determined by amino acid analysis and an bicinchoninic acid (BCA) assay described by Smith et al., Anal. Biochem., 150:76-85 (1985). Typically, several hundred milligrams of > 95% pure material was isolated from 100 g of starting material.

For small scale purifications, cells were grown in a 500 ml shaker flask containing 300 $\mu g/ml$ kanamycin until an absorbance of 0.6 at 600 nm was reached. Cells were then induced with IPTG for 3 hours at 37°C followed by centrifugation at 14,000 x g for 30 minutes. The cell 35 paste was resuspended in 20 ml of 1X phosphate buffered

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saline (PBS, Gibco) and sonicated for 3 minutes at 4°C using a Braun-Sonic model 1510 sonicator at 200 W. The lysed material was centrifuged for 30 minutes at 13,000 x g, and the precipitated material was resuspended in 5 buffer containing 6 M Guanidine-HCl as described above. Small scale purification of chemokine proteins was identical to the purification described above for the large scale isolation of chemokines.

Determination of Protein Concentration

Protein concentrations were determined using the BCA protein assay described above and results were calculated based on concentrations obtained from a standard dilution series of bovine serum albumin. Amino acid analysis of chemokines was obtained following 15 hydrolysis of the proteins for 24 hours at 90°C in 12 M HCl. Following hydrolysis, samples were derivatized and analyzed according to the method of Bidlingmeyer et al., J. Chromatography, 336:93-104 (1984).

Specific Chemokine-Like Proteins

The genes that express the new chemokine-like 20 proteins can be made by either of two methods. In the first method, a wild-type chemokine gene, e.g., one of the IL-8 or PF4 genes described above, is used as a template, and the desired amino acid substitutions are 25 made in this amino acid template by PCR amplification using, e.g., a sense 5' amplimer that contains the sequence for the desired modified codon(s). This results in the amplification of a new gene that encodes a new protein with the desired amino acid sequence.

Alternatively, genes encoding the new chemokinelike proteins can be synthesized without a template using the four overlapping amplimer strategy described above with amplimers containing appropriate sequence changes such that the final amplified DNA fragment will encode

the desired amino acid sequence in the expressed new chemokine-like protein.

A variety of new chemokine-like proteins were made using the wild-type amino acid sequences of human PF4 and 5 IL-8 (Figs. 3A and 4A, respectively) as the template. As shown in Fig. 2, these new proteins are similar in amino acid sequence to certain wild-type chemokines, in this case IL-8 or PF4, but include specific amino acid substitutions and/or insertions that make these new 10 proteins distinct and unique compared to the wild-type chemokines. In addition, these substitutions provide the new chemokine-like proteins with an unexpected enhanced myelosuppressive activity.

In particular, as shown in Fig. 4B, the cDNA (SEQ 15 ID NO:41) encoding new protein IL-8M1 (SEQ ID NO:70) was synthesized using a 5' sense amplimer IL-8M1-1 (SEQ ID NO:42), which contains a nucleotide mutation designed to result in the substitution of the amino acid sequence DLQ (an active domain from PF4) for the sequence ELR in wild-20 type IL-8 in the amino terminus of the protein. Because the remainder of the IL-8M1 sequence was to be the same as the wild-type IL-8 sequence, the 3' antisense amplimer used was the same one used in the synthesis of the cDNA encoding wild-type IL-8 described above, amplimer IL-8-4 (SEQ ID NO:40). This amplimer was chosen because no other mutation was desired, and this amplimer will produce no further mutation in the wild-type IL-8 sequence.

The cDNA (SEQ ID NO:50) encoding the new protein 30 IL-8M8 (SEQ ID NO:63) was synthesized as described above, except that the primer IL-M8-1 (SEQ ID NO:53) was used as the 5' amplimer. As shown in Fig. 4C, the IL-8M8-8 (SEQ ID NO:52) amplimer contains a nucleotide mutation designed to result in the substitution of the amino acid sequence $\rm E_4L_5Q_6$ for the sequence $\rm E_4L_5Q_6$ in the N-terminus

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of wild-type IL-8. Because the remainder of the IL-8M8 sequence was to be the same as the wild-type IL-8 sequence, the 3' antisense amplimer used was the same one used in the synthesis of the CDNA encoding wild-type IL-8 described above, amplimer IL-8-4 (SEQ ID NO:40).

The cDNA (SEQ ID NO:54) encoding the new protein IL-8M9 (SEQ ID NO:64) was synthesized as described above, except that the primer IL-8M9-1 (SEQ ID NO:55) was used as the 5' amplimer. As shown in Fig. 4D, the IL-8M9-1 (SEQ ID NO:55) amplimer contains a nucleotide mutation designed to result in the substitution of the amino acid sequence D₄L₅R₆ for the sequence E₄L₅R₆ in the N-terminus of wild-type IL-8. Because the remainder of the IL-8M9

sequence was to be the same as the wild-type IL-8

15 sequence, the 3' antisense amplimer used was the same one
used in the synthesis of the cDNA encoding wild-type IL-8

described above, amplimer IL-8-4 (SEQ ID NO:40).

The cDNA (SEQ ID NO:50) encoding the new protein

II.-8M10 (SEQ ID No:65) was synthesized as described

20 above, except that the primer IL-8M10-1 (SEQ ID NO:51)

was used as the 5' amplimer. As shown in Fig. 4E, the

II.-8M10-1 (SEQ ID NO:51) amplimer contains a nucleotide

mutation designed to result in the substitution of the

amino acid sequence D₄L₅N₆ for the sequence E₄L₅R₆ in the

25 N-terminus of wild-type IL-8. Because the remainder of

the II.-8M10 sequence was to be the same as the wild-type

II.-8 sequence, the 3' antisense amplimer used was the

same one used in the synthesis of the cDNA encoding

wild-type IL-8 described above, amplimer IL-8-4 (SEQ ID

30 NO:40).

The cDNA encoding IL-8M3 (SEQ ID NO:56) was synthesized in a manner similar to that described above for IL-8M1, IL-8M8, IL-8M9, and IL-8M10, except that the PCR amplification was designed to insert the amino acid sequence $E_{24}Y_{25}R_{26}R_{27}$ for the wild-type IL-8 sequence

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 $\rm E_{24} \rm L_{25} \rm R_{26} \rm V_{27}$. The nucleotide sequence encoding IL-8M3 (SEQ $^{-}$ ID NO:56) is shown in Fig. 4F.

As shown in Fig. 3B, the cDNA (SEQ ID NO:34) encoding new protein PF4M2 (SEQ ID NO:67) was synthesized using a 5' sense amplimer PF4M2-1 (SEQ ID NO:35), which contains a nucleotide mutation designed to result in the substitution of the amino acid sequence MSAKELRCQC (SEQ ID NO:45) for the sequence EAEEDGDLQCLQ (SEQ ID NO:40) in wild-type PF4 in the amino terminus of the protein. The second amplimer used was the same one used in the synthesis of the cDNA encoding wild-type PF4 described above, the 3' antisense amplimer PF4-4 (SEQ ID NO:33).

The chemokines PF4-414 and PF4-426 were synthesized using a similar techniques. The synthesis of PF4-414 is illustrated in Fig. 3c, using the amplimers for wild-type PF4 and substitution the amplimer PF414-1 (SEQ ID NO:58) for the antisense 3' amplimer. The synthesis of PF4-426 (SEQ ID NO:68) is illustrated in Fig. 3D using amplimers PF4-426-1 (SEQ ID NO:60), PF4-26-2 (SEQ ID NO:61), PF4-426-3 (SEQ ID NO:62), and the amplimer PF4-4 (SEQ ID NO:33).

Pased on the methods, techniques, and assay results described herein, additional new proteins can be created using other wild-type chemokines as the template or structural basis. As described below and shown schematically in Fig. 5, the amino acid changes in a template based on a first wild-type chemokine must introduce at least one new myelosuppressive active domain from one region of a second wild-type chemokine into the template, while maintaining a myelosuppressive active domain of the first chemokine in another region of the template or introducing another active domain from the second or a third wild-type chemokine.

Fig. 5 shows that wild-type IL-8, having active 35 domains 1 (ELR of IL-8) and 2 (ELRV of IL-8), provides a

normal level of myelosuppressive activity. Wild-type PF4, having active domains 3 (the first DLQ of PF4) and 4 (the second DLQ of PF4), also provides a normal level of myelosuppressive activity. If either of these active domains are removed, the resulting protein loses its myelosuppressive activity. For example, PF4M1, which is missing domain 3; PF4-412, which is missing domain 4; and IL-8M4, M3, and M6, which are all missing domain 2; all lack myelosuppressive activity. On the other hand, new

10 chemokine-like protein PF4M2 includes active domains 1 and 4, and has unexpectedly high myelosuppressive activity. Similarly, IL-8M1 includes domains 3 and 2, and has unexpectedly high myelosuppressive activity.

Framples of new chemokine-like proteins based on

Examples of New Chemother The Provided Provided

20 (PF4's first active domain) proximal to and on the Nterminus side of the first cysteine to the amino acids DIR. As shown by the assay results described below, this change destroyed PF4's first active domain and resulted in a protein without myelosuppressive activity.

25 Testing of PF4M2 showed that replacing an active domain in PF4 with an active domain from a second chemokine creates a new highly myelosuppressive protein. PF4M2 was generated by replacing the entire N-terminus up to the first cysteine of wild-type PF4 with the 30 corresponding N-terminus of wild-type IL-8. This change

corresponding N-terminus of wild-type IL-8. This change replaced PF4's first active domain with IL-8's first active domain ($E_4L_5R_6$) while maintaining PF4's second active domain ($D_54L_55Q_56$), and resulted in a highly active new protein.

New proteins PF4-412, PF4-413, and PF4-414 were generated by replacing amino acids 51 to 61 of wild-type PF4 with the corresponding amino acids from wild-type IL-8, NAP-2, or GROG, respectively. Of these new proteins, 5 PF4-412 and PF4-413 were essentially inactive, because the substitution of the new domains from IL-8 and NAP-2 destroyed PF4's second active domain without introducing a new active domain. PF4-414 retained the first active domain of PF4 and included an active domain from GROG, and had a higher activity than wild-type PF4.

PF4-421 was generated from an exchange of amino acids 25 to 31 of wild-type PF4 with the corresponding amino acids from wild-type IL-8. This resulted in a new protein that maintained PF4's two active domains and introduced IL-8's second active domain (E₂₄L₂₅R₂₆V₂₇). This protein had about the same activity as wild-type PF4.

pF4-426 was created by replacing all three Arg (R) residues of wild-type PF4 with Glu residues (Q) to 20 achieve a novel region at amino acids 42 to 50. These changes created a region in the new protein equivalent to a corresponding region in GROβ that includes an active domain, while maintaining the first active domain of PF4. As a result, PF4-426 was highly active.

Examples of new chemokine-like proteins based on IL-8 include IL-8M1, IL-8M3, IL-8M4, IL-8M6, IL-8M7, IL-8M8, IL-8M9, IL-8M10, and IL-8M64. IL-8M1 was generated by replacing the amino acids ELR proximal and on the N-terminus side of the first cysteine of normal 30 IL-8 (IL-8's first active domain) with the amino acids DLQ from the corresponding location in wild-type PF4. Thus, this new protein included PF4's first active domain and IL-8's second active domain and was highly active.

IL-8M3, IL-8M4, and IL-8M6 were generated by 35 creating one or two point mutations within the IL-8 dimer

interface (IL-8's second active domain including the amino acids ELRV between the second and third cysteines in normal IL-8). These mutations destroyed IL-8's second active domain and, as expected, resulted in inactive

5 proteins.

IL-8M7 was generated by replacing the entire C-terminus after the fourth cysteine of normal IL-8 with the corresponding C-terminus of normal PF4. This substitution resulted in a new protein having IL-8's 10 first and second active domains and PF4's second active domain, and had about the same activity as wild-type IL-8.

IL-8M8, IL-8M9, and IL-8M10 were generated by creating one or two point mutations within the DNA 15 encoding the amino acids ELR proximal and on the Nterminus side of the first cysteine of normal IL-8 (IL-8's first active domain). The IL-8M10 protein $(D_4 L_5 N_6)$ exhibited myelosuppressive activity that was significantly enhanced relative to wild-type IL-8 20 activity, and comparable to the myelosuppressvie activity of IL-8M1. The mutations in IL-8M8 and IL-8M9 ($E_4L_5Q_6$ and $\mathrm{D_4L_5R_6}$, respectively) resulted in proteins having myelosuppressive activities comparable to that of IL-8 and PF-4, suggesting that the double mutation within IL-8 25 is critical to the enhancement of myelosuppressive activity. These results also indicate that the amino acid sequences DLN, DLQ, and ELQ can also serve as "active domains," and can, in combination with another active domain, confer myelosuppressive activity upon the 30 novel chemokine-like protein.

IL-8M64 was generated by truncating the last eight amino acids from the C-terminus of normal IL-8, which resulted in a new protein with about the same activity as wild-type IL-8.

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Testing of Specific New Chemokine-Like Proteins

The different chemokine-like proteins shown in Fig. 2 were tested in both in vitro and in vivo assays to demonstrate their ability to inhibit proliferation of 5 progenitor cells. Wild-type PF4 and IL-8 by themselves were shown to inhibit stem cell proliferation at concentrations as low as 25 ng/ml, whereas the new chemokine-like proteins were shown to have equivalent myelosuppressive activity at much lower concentrations.

New Chemokine-Like Proteins Inhibit Progenitor Cell Proliferation In Vitro

Chemokine-like proteins made as described herein can be tested by standard in vitro assays to determine whether they inhibit formation of colony and cluster formation of granulocyte macrophages (termed colony forming unit-granulocyte macrophages (CFU-GM)). Such assays are well known and representative assays are described in Gentile et al., U.S. Patents Nos. 5,149,544 and 5,294,544. In these assays, bone marrow or spleen cells are stimulated with, e.g., CSF, in an in vitro culture system. The inhibitory activity of the new chemokine-like protein is measured as the amount it decreases the CSF-stimulated colony and cluster formation.

25 In particular, the new proteins described above were tested for inhibition of GM colony formation as follows. LD cells were plated at a density of 5 X 10⁵ cells in 0.3% agar culture medium with 10% FBS (Hyclone, Logan, UT) for assessment of CFU-GM. CFU-GM colonies 30 (>40 cells/group) were stimulated by human rGM-CSF (100 U/ml) in combination with human rSIF (50 ng/ml). All colonies were tested in the absence or presence of different concentrations of new chemokine-like proteins to examine potency of inhibition of proliferation.

Colonies were scored after 14 days incubation at lowered (5%) $\rm O_2$ tension, and 5% $\rm CO_2$ in a humidified

environment in an ESPEC N₂-O₂-CO₂ incubator BNP-210 (Taoi - ESPEC Corp., South Plainfield, NJ). Three plates were scored per determination. The results are expressed in Table IA below as a mean percent change from control ± 1 5 S.E.M. for the number of experiments shown in experiments in which the control number of colonies per 1 x 10⁵ cells/ml ranged from 115 ± 5 (mean ± 1 S.E.M.) to 382 ± 10. The inhibition shown, while partial inhibition of the total colony formation, is 90 to 100% inhibition of the steel factor enhanced colony formation.

In Table IA, those data marked with a "b"
represent a significant change from control, p < 0.01
(Students t test), while those data marked with a "c"
represent a significant change from control, p < 0.05.

The in vitro results are also shown in the graphs of
Figs. 6A to 6G.

Table IA
Chemokine concentration (ng/ml)

	Chemokine 100	50	25	10	. 1	0.1	0.01 0.001
20	PF4 WT -41±3b PF4M1 -9±10	-40±2b	2+2	-6+1			
	PF4M2 -43 ± 2b	-44 ± 2^{b}	-42±3b	-46±2b	-47±2b	-42±5 ^b	-19±8° -3±2
	PF4-412 -6±4 PF4-413 -19±10°	-3±3 -14±16	2+2	-2±1	+1±1		
25	PF4-414 -45±1b	$-44 + 1^{b}$	-44 ± 2b	-43 ± 2 ^b	-42±5 ^b	-43 ± 1^{6}	$-34 \pm 8^{b} -3 \pm 3$
	PF4-421 -42±6 ^b PF4-426 -45±3 ^b	-40±4b	-29 ± 3°	-25±9°	-8±4 -45±3⁵	+1±1 -44±4b	-40±3b -20±1c
	PF4-420 -43=3						
	IL-8WT -42±2 ^b IL-8M1 -43±4 ^b	-43±2 ^b	-21±5b	-4±2 -46±2b	-3±6 -46±2 ^b	-46 ± 2 ^b	-40±1b -25±4b
30	IL-8M1 -43±4 IL-8M3 -8±3	-4±3	-1±2	-7±2			
	IL-8M4 -1±2	-1±2 -1±2	+1±1				
	IL-8M6 -2±2 IL-8M7 -43±4 ^b	-45 ± 2 ^b	-25±2°	-2±3	0±2		
	IL-8M64 -43 ± 1b	-43±2b	-27 ± 5 ^b	-10±7	-8±5	-5 ± 1	

35 As shown in Table IA, wild-type PF4 and IL-8 were effective to inhibit colony formation by human bone marrow CM progenitor cells at concentrations of at least

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25 ng/ml. At 10 ng/ml, neither one provided more than insignificant inhibition.

New chemokine-like protein PF4M1, in which the first active domain of PF4 was destroyed, was essentially inactive. PF4M2, however, which included the first active domain of wild-type IL-8 and the second active domain of wild-type PF4, had a much higher activity than wild-type PF4. Although PF4-412 and PF4-413 both retained PF4's first active domain, they were essentially inactive because they both lacked a second active domain.

0 inactive because they both lacked a second active domain. PF4-414, which retained PF4's two active domains and included a new active domain from GROα, had a higher activity than wild-type PF4. PF4-421, which retained PF4's two active domains and included the second active

15 domain from wild-type IL-8, had a somewhat higher activity than wild-type PF4. PF4-426, which retained PF4's first active domain and included an active domain from wild-type GRO\$, was highly active.

New chemokine-like protein IL-8M1, which included 20 PF4's first active domain and IL-8's second active domain, was active down to 0.001 ng/ml. On the other hand, new proteins IL-8M3, IL-8M4, and IL-8M6 were essentially inactive, because each lacked an intact second active domain. IL-8M7, which included wild-type IL-8's first and second active domains and PF4's second active domain, had about the same activity as the wild-type IL-8. IL-8M6, which retained wild-type IL-8's first and second active sites, but which lacked the last

30 activity as wild-type IL-8.

The data in Table IA are also represented in graphical form in Figs. 6A to 6G. Fig. 6A shows the percent inhibition of myeloid progenitor colony formation by FF4M2 and FF4-413 compared to wild-type FF4 (PF4 WT).

8 amino acids of wild-type IL-8, also had about the same

35 Fig. 6B shows the percent inhibition by PF4-426 and PF4-412 compared to PF4 WT. Fig. 6C shows the percent inhibition by PF4-414, PF4-421, and PF4M1. Fig. 6D shows the percent inhibition by IL-8M1 and IL-8M3 compared to IL-8 WT. Fig. 6E shows the percent inhibition by IL-8M1,

40 IL-8M64, and IL-8M6. Fig. 6F shows the percent inhibition by IL-8M1, IL-8M7, and IL-8M4. Fig. 6G shows

the free transfer and

the percent inhibition by wild-type PF4, PF4-414, PF4-426, PF4M2, and wild-type IL-8. These graphs are all on the same scale, therefore, the activity of different new proteins can be directly compared.

The relative activities of IL-8M1, IL-8M8, IL-8M9, IL-8M10, wild-type IL-8, and wild-type PF4 in myelosuppression of colony formation by granulocyte macrophages (GM) (CFU-GM), erythroid cells (BFU-E), and multipotential cells (CFU-GEMM) were tested in an in

10 <u>vitro</u> assay similar to the assay described above (Lord et al., <u>Blood</u>, <u>79</u>:2605-2609, 1992); and Maze et al., <u>J. Immunol.</u>, <u>149</u>:1004-1009, 1992).

Briefly, the effects of chemokines upon CFU-GM were assessed by plating 1 x 10⁵ low density

15 (<1.077 gm/cm³) normal human bone marrow cells in 0.3[‡] agar culture medium with 10[‡] FBS (Hyclone, Logan, UT) with 100 U/ml recombinant human (rhu) GM-CSF plus 50 ng/ml rhu Steel factor (Inmunex Corp., Seattle, WÅ). The effects of chemokines upon CFU-GEMM and BFU-E were assessed by growing the cells in 0.9[‡] methylcellulose culture medium in the presence of recombinant human erythropoietin (rhuEpo) (1 to 2 U/ml) in combination with 50 ng/ml rhu Steel factor. Three plates were scored per concentration per experiment for CFU-GM, CFU-GEMM, and 25 BFU-E colonies after incubation at 37°C in lowered (5[‡]) O, for 14 days. The combination of GM-CSF and Steel

which are derived from early, more immature subsets of CFU-GM, CFU-GEMM, and BFU-E. Levels of significance were determined using Student's t-distribution (two-tailed test). The data are presented in Table IB, with "a" denoting a significant change from control with p<0.01, and "b" denoting a significant change from control at

factor, or erythropoietin and Steel factor, allowed detection of large colonies (usually >1,000 cells/colony)

35 p<0.05 (student's t test).

As shown in Table IB, there were no observable differences in the myelosuppressive activities of the chemokines between cell lineages. These data indicate that chemokine-dependent suppression is not cell lineage-

- 5 specific, but rather affects multiple progenitor cell populations. IL-8M8 and IL-8M9 exhibited myelosuppressive activities comparable to the myelosuppressive activities of wild-type IL-8 and PF4 wild-type. IL-8M10 was highly active in
- 10 myelosuppression, and had activity comparable to that of IL-8M1, suggesting that replacement of Arg₆ of wild-type IL-8 with either glutamine or asparagine can mediate enhanced myelosuppressive activity if aspartic acid is present at position 4 of the IL-8 wild-type sequence.

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					Tab	Table IB				
_	Other Part and			פ	nemokine	Chemokine Concentration (ng/ml)	ion (ng/	1)		
	CHEMONALIE	100	50	25	10	1	0.1	0.01	0.001	0.0001
	NO-HAV									
	11 0 11	-48+6ª	-49+58	-26+5ª	-5±3					
٠.	10-0 41	-404E	-49+6ª	-34+7ª	-8+4	-3±1				
	FF4 WI	-E1+48	-51+58	-50+5ª	-49+4ª	-49±5ª	-47±5ª	-48±3ª	-28±3ª	-7±3
	TL-OMI	-53+4ª	-51+4ª	-34+5ª	-18±8b	-13±8	-4+2			
	owo TT	BA463-	-50+4ª	-32+7ª	-17±10	-16±8	-3±2			
	TI8M10	-54+4g	-52±3ª	-51±5ª	-49+4ª	-54±4ª	-49+3ª	-48±1ª	-32±5ª	-3+4
_	a-mau									
	E 0 12	-5146a	-48+8ª	-21+6 ^b	9 + 2+	+2+7				
	TH 0-17	17.74	┰	-23+17	-2±5	-5±2				
	11 1001	- FO+1	┰	-50+1ª	-46±1ª	-48±2ª	-46±5ª	-45+2ª	-14±6	-4+9
	TOUL I	107	+-	-29+4 ^b	-1+2	-7±1				
	TI-8M8	CT64-	-	-	-10+6	-6+3				
12	IL-8M9	484	-	4-	-47+78	-43+7ª	-43+7ª	-38±7ª	-22±5b	+2+5
	IL-8M10	-517-	-30±2	-1∥						
	CFU-GEMM									L
	IL-8 WT	-66±7ª	-64±0ª	-	-7+2	0+0		-		
	PF4 WI	-64±0ª	-62±7ª	-27±9b	-7±2	0+0			1	1
20	IL-8M1	-64±0ª	-66±1ª	$\overline{}$	\dashv	-62±3ª	-62±3"	-55+0"	-36±0	-177
	IL-8M8	-64±0ª	-64±0ª	_	-27±18	-5+5				1
	TI,-8M9	-64+0a	-64±0ª	-32±14 ^b	-18±18	-6+3			1	
	TT-8M10	-66+2ª	-65+2ª	-64±0ª	-63±3ª	-64±0ª	-60±5ª	-50+5	-30+72	-5+5
				ł						

The data in Table IB are also represented in graphical form in Fig. 6H, which shows the percent inhibition of GM colony formation by IL-8M1 and IL-8M10 compared to wild-type IL-8 (IL-8 WT) wild-type PF4 (PF4 WT).

As noted above, all of the results, taken together, show that there are two active domains within IL-8 involved in myelosuppressive activity (E₄L₅R₆ and E₂₄L₂₅R₂εV₂₇). Moreover, the amino acid sequences DLN (IL-10 8M10), ELQ (IL-8M8), or DLR (IL-8M9) can also function as active domains: Likewise, PF4 also has two active domains (D₁L₆C₉ and D₅₄L₅₅C₅₆). Furthermore, the data obtained from PF4-426 shows that the amino acid sequence IATLKNGQK (SEQ ID NO:43) from wild-type Gro-β is responsible, in conjunction with PF4's first active domain, for the high myelosuppressive activity observed. This sequence, which is present in Gro-β, but not in Gro-α or Gro-γ, is therefore believed to contain an active domain of this protein.

In addition, the data obtained from PF4-414 shows 20 that the amino acid sequence ACLNPASPIVK (SEQ ID NO:44) from wild-type $Gro-\alpha$ is responsible, in conjunction with PF4's first active domain, for a high myelosuppressive activity. Although $Gro-\alpha$ is itself inactive in 25 suppressing myeloid cell proliferation, it competes with IL-8 and PF4 for progenitor cell suppression, which suggests that it is able to interact on the cell surface and block activity. Thus, it is possible that $Gro-\alpha$ lacks some activation sequence required for 30 myelosuppression, but includes a sequence responsible for binding. Without the activation sequence, wild-type Gro- α has no myelosuppressive activity, but remains a competitive binding protein. However, when combined with an active domain from PF4 as described herein, the 35 resulting new protein has very potent myelosuppressive

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activity. This sequence ACLNPASPIVK (SEQ ID NO:44) is therefore believed to contain an active domain of $Gro-\alpha$.

New Chemokine-Like Proteins That Inhibit Myelosuppression Do Not Elicit Neutrophil

5 Degranulation

The ability of the new chemokine-like proteins to mediate neutrophil degranulation was tested in an in vitro assay as described by Hebert, et al., J. Immunol., 145:3033, 1990, with some modifications. Isolated human neutrophils were suspended in PBS containing 0.02 M Na₂HPO₄, pH 7.4, 0.15 M NaCl, 0.2 M Hepes, 1 mg/ml BSA, 5 mM glucose, 5 x 10⁻³ mg/ml cytochalasin B (5 mg/ml stock in DMSO; Sigma) at a concentration of 2 x 10⁶ cells/ml. 0.5 ml aliquots of the resuspended neutrophils were added to 0.5 ml of the PBS solution. Following incubation, the cells were reequilibrated at 37°C for 15 min.

Varying concentrations of wild-type IL-8 or IL-8 polypeptide antagonist were added to the neutrophils with gentle mixing. Following addition of the IL-8 or IL-8 20 polypeptide antagonist, the cells were pelleted and 0.75 ml of the resulting supernatant was added to 2.25 ml PBS in the presence of 5 x 10⁻³ mg/ml of the fluorescent elastase substrate

MeO-Suc-Ala-Ala-Pro-Val-aminomethylcoumarin (5 mg/ml

- stock in DMSO; Peninsula Laboratories, Inc., Belmont, ch). Cleavage of the fluorescent elastase substrate releases a molecule which, upon excitation at 380 nm, emits light at a wavelength of 460 nm. The samples were incubated for 1 h at 37°C, placed on ice, and analyzed
- 30 using a spectrofluorometer. Elastase levels were determined as the amount of fluorescent probe released from the elastase substrate. Uncleaved substrate displayed minimal fluorescence intensity relative to the cleaved product. Backgrounds, which were typically less
- 35 than 2% of the maximum fluorescence released at the highest chemokine concentration were subtracted from each

ment to the state of the

sample. The activities of compounds in this in vitro neutrophil degranulation assay are predictive of the ability of the compounds to elicit neutrophil degranulation in vivo.

- 5 As shown in Fig. 7A, the antagonist, IL-8M1 was significantly inhibited in its ability to activate neutrophils, which as quite surprising given this protein's otent myelosuppressive activity. Without being held to theory, the D₇L₈Q₉ amino acids (first active 10 domain) from wild-type PF4 appear to make this new chemokine-like protein act as if PF4 and IL-8 were both present and acting synergistically, but with a much higher activity per total amount of prc ein than can be achieved by merely combining the two wild-type proteins.
- 15 With the exception of PF4M2, which possesses the ELR domain of IL-8, none of the new chemokine-like proteins based on PF4, including the highly active PF4-414 and PF4-426, elicited any response on human neutrophils (Fig. 7A). New chemokine-like proteins IL-8M1, IL-8M8, 20 IL-8M9, and IL-8M10, which are able to suppress
 - proliferation of myeloid progenitor cells, elicited no or little neutrophil degranulation relative to wild-type IL-8 (Fig. 7B). Since IL-8M1 and IL-8M10 are highly active at
- 25 inhibiting proliferation of progenitor cells, yet do not activate neutrophils, these new chemokine-like proteins will not elicit the adverse neutrophil activation and concomitant inflammatory effects observed with wild-type II-8.
- 30 New Chemokine-Like Proteins Inhibit Progenitor Cell Proliferation In Vivo

Wild-type PF4 and IL-8, and new chemokine-like protein IL-8M1 were also tested in an <u>in vivo</u> model as follows. Murine models for assessing progenitor cell proliferation were carried out substantially as described

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in Cooper et al., Exp. Hematol., 22:186-193 (1994). The results of this in <u>vivo</u> model, together with the <u>in vitro</u> assay results described above, are predictive of the efficacy of the new proteins in treating patients, e.g.,

5 humans.

Control medium, control diluent, wild-type IL-8
and PF4, and the new chemokine-like protein IL-8M1 were
evaluated for effects on myelopoiesis in vivo in mice,
with endpoints being nucleated cellularity and

10 differentials in the bone marrow, spleen, and peripheral blood, and absolute numbers and cycling status of myeloid progenitor cells in the marrow and spleen. In each test, groups of three C3H/HeJ mice or BDF₁ mice were exposed to a particular test sample. C3H/HeJ mice were used because 15 they are relatively insensitive to the effects of endotoxin. Thus, any potential endotoxin contamination in the chemokine samples did not influence the in vivo results. BDF₁ mice were used in addition to C3H/H3J mice to make certain that the observed phenomena were not

C3H/HeJ and BDF₁ mice were purchased from the Jackson Laboratory (Bar Harbor, ME), were housed in a conventional animal facility, and were injected intravenously with 0.2 ml/mouse sterile pyrogen-free 25 saline, or the stated amount of a specific chemokine or chemokine-like protein, as described in Mantel et al., P.N.A.S., USA, 90:2232 (1993). The mice were sacrificed 24 hours later.

The cycling status of hematopoietic progenitor

30 cells, i.e., the proportion of progenitor cells in DNA
synthesis (S phase of the cell cycle), was estimated as
described in Maze et al., J. Immunol., 149:1004 (1992)
and Cooper et al., Exp. Hematol., 22:186 (1994). The
high specific activity (20 Ci/mM)-tritiated thymidine (50
35 µCi/mL) (New England Nuclear, Boston, MA) kill technique

was used, and is based on a calculation in vitro of the reduction in the number of colonies formed after pulse exposure of cells for 20 minutes to "hot" tritiated thymidine as compared with a control such as McCoy's medium or a comparable amount of non-radioactive "cold" thymidine.

In brief, femoral bone marrow was removed from the sacrificed mice, treated with high-specific-activity tritiated thymidine, and plated in 0.3% agar culture 10 medium with 10% FBS in the presence of 10% volume/volume pokeweed mitogen mouse spleen cell cultured medium. Colonies (>40 cells/aggregate) and clusters (3-40 cells) were scored after 7 days of incubation.

Three plates were scored for each sample for a 15 statistical analysis. Each mouse was evaluated separately in groups of three mice each. Results for two experiments with three mice per group (each analyzed individually) are shown in Table II below, and are expressed as the mean \pm 1 standard deviation (SD). The 20 results are derived from the averages of each of the individual mice within a group. The probability of significant differences between groups was determined by Student's t-test (two-tailed). Parentheses indicate the percent change from control. In the table, those data 25 marked with a "C" represent a significant percent change from control, p < 0.005 (Students t test), while those data marked with a "d" represent a significant percent change from control, p < 0.025, and those data marked with an "" represent a significant percent change from 30 control, p<0.01. All other differences are not significantly different from control, p > 0.1.

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Table II

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Influence of IL-8, IL-8M1, and PF4 on Absolute Numbers and Cycling Status of Granulocyte Assembles Propention Cells (CFU-GM) in Vivo in Femure of C3H/He1 and BDF, Mice.

	-Macrophagerrog	emilor cens (cr.o.	-Macrophagerrogenilor Cells (Cr.OO.M.) III 1 Cilians of Constitution	10 611		
	nucleated cells/Femur x 10 ⁻⁶		Absolute No. CFU - GM/Femurx 10-3	J-GM/Femurx	Percent CFU - GMin Cycle	GMin Cycle
	BDF,	C3H/HeJ	BDF ₁	C3H/H6J	BDF ₁	C3H/HeJ
control saline	133±1.6	25.7±2.5	9.1±0.5	28.1±3.7	57.0±6.8	55.0±5.1
ILSM 10se	13.3+1.8	232±4.7	6.1±0.8 (-33) ^c	14.6±1.6 (-48.1) ^C	32±64 (-94) ⁶	05±2.9 (-99) ^c
II.8M lue	10.0+1.0	213±1.4	b(21-) 20±1.77	263+3.9 (-6.3)	48.9±3.3 (-14)	485±87 (-12)
11_8M 001ue	10.7+0.7	23.7±0.4	B.1±1.1 (-12)	295±2.2 (+4.8)	49.0±1.8 (-14)	() 65 + 915
196 196	115+20	23.0+3.9	S.1±0.3 (+4) [€]	16.2±2.4 (42.4) ⁶	0.7±45 (-99) ^c	22 <u>+</u> 43 (-96) ^c
TOWN THE	12/0+27	22.1+2.4	5.6±05 (-38) ^c	15.6±0.6 (-44.7) ^C	35±7.7 (-94) ^c	29±3.0 (-95) ^c
ILAMI OOLur	123+19	23.1±5.4	6.7±0.8 (-26) ⁶	16.9 <u>+</u> 4.2 (-39.7) ^d	17.7±5.3 (-69) [¢]	14.7±11.7 (-73) ^C
10.4	126+10	25.0±6.7	6.1±0.3 (-33) ^c	17.1±4.2 (-39.3) ^d	3.3±5.6 (-94) ^C	15±61 (-97) ^C
9001 730	124+26	21.6±1.0	8.8±1.7 (-3)	29.7±2.4 (+5.5)	52.0±6.8 (-9)	50.9±7.6 (-8)
PF4 0.01ug	12.1±1.0	21.8±2.1	8.6±0.4 (-5)	268±34 (48)	48.6±1.9 (-15)	50.8±5.8 (-8)

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The in vivo results are shown in Table II above (and are represented in graphical form in Figs. 8A and 8B). As shown, wild-type IL-8 and PF4 were effective at a dosage of 10 $\mu g/mouse$, but were not significantly 5 effective at lower dosages of 1.0 or 0.01 $\mu g/mouse$. Similarly, cell cycling was completely inhibited at a dosage of 10 μ g/mouse. As expected, these in vivo data confirmed the results of the in vitro assay described above.

The new chemokine-like protein IL-8M1, however, 10 was effective at a dosage as low as 0.01 $\mu g/mouse$. At this dosage, progenitor cell cycling was suppressed from approximately 55% down to 14.7% +/- 11.7%, for a total reduction of 73.3% compared to control. The results were 15 similar for the two studies of absolute number of colony forming units per mouse femur (top of Table II), and the percentage of cells in the replicating cycle (bottom of Table II).

The results of the in vivo tests are also depicted 20 in graphical form in Figs. 8A and 8B. As shown in Fig. 8A, dosages of 1.0 and 0.01 μg of wild-type IL-8 and PF4 had little effect on the percentage of CFU-GM in cycle, which is normally about 50 to 55 percent. However, even at a dosage of 0.01 μ g, new chemokine-like protein IL-8M1 25 reduced the percentage of cells in cycle to less than 15 percent, and to less than 5.0 percent at a dosage of 1.0 μ q. IL-8M1 also reduced the percentage of cells in cycle to less than 5.0 percent at a dosage of 10.0 μg , as did wild-type IL-8 and PF4.

As shown in Fig. 8B, absolute numbers of CFU-GM progenitor cells were also affected by the presence of chemokines. Wild-type PF4 and IL-8 decreased total cell numbers only at the 10 μg doses, while IL-8M1, being the more potent agent, inhibited the proliferation of 35 progenitor cells at doses as low as 0.01 $\mu g/mouse$. The

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decrease in absolute numbers of progenitor cell in vivo indicates that cell proliferation has been inhibited, and is a good correlate of the cell cycling assay described above.

Binding of New Chemokine-Like Proteins to

5 IL-8 Receptor Subtype B In Vitro The ability of the IL-8 derived chemokines to competitively bind IL-8 receptor subtype B in the presence of wild-type IL-8 was examined in vitro. Briefly, IL-8 was iodinated as previously 10 described (Thomas et al., J. Biol. Chem., 266:14839-14841, 1991), or purchased from NEN-Dupont. A stable transfectant CHO cell line, 4ABCH033 (LaRosa et al., <u>J. Biol. Chem., 267</u>:25402-25406, 1992), expressing human 15 neutrophil IL-8 receptor subtype B (huIL8Rb), was used in binding assays to test mutant chemokine binding. Approximately 5 \times 10³ cells were plated in the wells of a 96 well plate and grown for 2 to 3 days. The medium was replaced with phosphate-buffered saline (PBS) 20 supplemented with 0.5% BSA (PBS/BSA), 0.5 nM ^{125}I -labeled IL-8, and increasing concentrations of the various unlabeled chemokines. The plates were incubated on ice for 90 minutes, the binding mixture was removed, and the cells were washed three times with ice-cold PBS/BSA. The 25 cells were removed from the plates with trypsin-EDTA (Gibco), and the radioactivity of the entire contents of the well determined by gamma counting. Binding of each chemokine to untransfected cells, or binding in the presence of 2 μM unlabeled IL-8, was determined and the 30 value obtained used as the background value. Binding data was plotted as percent of mean specific binding (total binding minus background/specific binding with no competitor). The Ki (the chemokine concentration to inhibit 50% of 125 I-labeled wild-type IL-8 binding) for

35 each chemokine was calculated and is presented in Table III. "NC" indicates no competition.

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Table III

Chemokine	Ki_(nM)
IL-8	3.0 <u>+</u> 0.5
IL-8M1	661.0 <u>+</u> 189
IL-8M3	22.1 <u>+</u> 4.4
IL-8M4	29.3 <u>+</u> 7.2
IL-8M64	35.3 <u>±</u> 6.2
PF4	NC
P.F4M1	839.4 <u>+</u> 202
PF4M2	52.0 <u>+</u> 11.8

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Wild-type IL-8, as well as other "ELR"-containing chemokines including Gro-α, Gro-β, and NAP-2, bound the IL-8 subtype B receptor with high affinity. As shown in Table III, competition binding experiments utilizing

15 1251-labeled IL-8 and unlabeled mutant chemokine

- competitors demonstrated that the IL-8 receptor subtype B binding and myelosuppressive activity are not correlated. For example, IL-8M1 and PF4-M2 have comparable myelosuppressive activity, yet neither chemokine
- 20 effectively competes with wild-type IL-8 for IL-8 receptor subtype B binding. Moreover, the chemokines that most effectively competed with wild-type IL-8 for IL-8 receptor subtype B binding exhibited the lowest levels of myelosuppressive activity (e.g., IL-8M3,
- 25 IL-8M4, and IL-8M64). These data suggest that myelosuppression is not mediated by chemokine binding to the IL-8 receptor subtype B.

Heparin Binding by PF4 and PF4-based Chemokines
In addition to its myelosuppressive activities,
30 wild-type PF4 is characterized by high affinity binding
and neutralization of heparin. Wild-type PF4 binds
heparin with a binding constant between 3 x 10⁻⁸ and 1 x
10⁻⁷ M. The heparin binding affinity for the PF4-derived

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chemokines was determined according to methods well known in the art. The PF4-derived chemokines bound heparin with a dissociation constants approximating 1 x 10⁻⁷ M, suggesting that the mutant proteins, including PF4-426, had properly refolded and bound heparin in a manner analogous to wild-type PF4. All of the IL-8 related proteins, including wild-type IL-8, bound heparin at levels at least 20-fold lower than PF4, suggesting that the IL-8-derived chemokines were also folded in the 10 proper conformation.

Examination of Chemokine-like Competitors of Myelosuppression

Chemokine-like proteins that were inactive in the progenitor proliferation assays above were tested for 15 their ability to inhibit the myelosuppressive activity of either IL-8 or PF4. Varying concentrations of chemokine mutants IL-8M3, IL-8M4, IL-8M6, and PF4-412 were incubated with either 50 ng/ml wild-type PF4 or 50 ng/ml wild-type IL-8 in the CFU-GM assay described 20 above. Cells were plated in 0.3% agar culture medium with 10% v/v fetal bovine serum, 100 U/ml rhu GM-CSF, and 50 ng/ml rhu steel factor. The results of these experiments, as mean \pm 1 standard error of the mean (SEM) with percent change from control shown in parenthesis, 25 are shown in Table IV. These results are one of two similar and reproducible experiments. "b" indicates significant change from control at p<0.001; "c" indicates significant change from control at p<0.05.

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				able IV				
	Sample	Concentration		-	Wild-	type _L-8	Wild-tyr	e PF4
	control	medium	62 <u>±</u> 1		31 <u>+</u> 4	(-50) ^b	32 <u>+</u> 3	(-48) ^b
	IL-8M3	500	60±3	(-3)	60 <u>+</u> 5	(-3)	33 <u>+</u> 1	(-47)b
5	IL-8M3	250	58 <u>+</u> 2	(-3)	63 <u>+</u> 2	(+2)	28 <u>+</u> 2	(-55)b
	IL-8M3	50	62 <u>+</u> 4	(0)	26 <u>+</u> 1	(-58) ^b	26 <u>+</u> 2	(-58)b
	IL-8M4	500	61 <u>+</u> 2	(-2)	33 <u>+</u> 2	(-47)b	29 <u>+</u> 3	(-53) ^b
	IL-8M4	250	61 <u>+</u> 1	(-2)	34 <u>+</u> 1	(-45)b	29 <u>+</u> 2	(-53) ^b
	IL-8M6	500	32 <u>+</u> 4	(-48) ^b				
10	IL-8M6	250	47±4	(-24)°				
	IL-8M6	50	61 <u>+</u> 2	(-2)				
	PF4	500	57 <u>±</u> 3	(-8)	27 <u>+</u> 2	(-56) ^b	27 <u>+</u> 2 (-56) ^b	

Mutants IL-8M4 and PF4-412 had no effect either as suppressors of progenitor proliferation, or as
15 competitors with either wild-type IL-8 or wild-type PF4.
IL-8M6 suppressed progenitor cell proliferation at the highest concentration tested (500 ng/ml), thus demonstrating its activity as a weak agonist in this system.

- 20 IL-8M3 did not exhibit suppression of progenitor cell proliferation at any concentration tested. However, at 250 ng/ml and 500 ng/ml, IL-8M3 blocked the myelosuppressive activity of wild-type IL-8. Under identical conditions, this mutant was unable to inhibit
- 25 the ability of PF4 to suppress progenitor proliferation. Thus, IL-8M3 can enhance progenitor cell proliferation by inhibiting IL-8 mediated myelosuppression.

IL-8M3 inhibition of IL-8-mediated myelosuppression also suggests that this mutant interacts 30 with a cell-based receptor. The data from the receptor binding studies above, which indicate that IL-8M3 competes with wild-type for IL-8 receptor subtype B binding, suggests that IL-8M3 mediates its anti-

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myelosuppressive activity by binding this receptor, or a similar, IL-8 subtype B-like receptor. In addition, the inability of IL-8M3 to inhibit PF4-mediated myelosuppression suggests that several different progenitor cell receptors, with different specificities, facilitate progenitor cell myelosuppression.

Uses of New Chemokine-Like Proteins

The new chemokine-like proteins can be used both as diagnostic and therapeutic agents.

10 Diagnostic Uses

The subclass of progenitor cells that respond to the presence of chemokines has been shown to include a cell population which contains the cell surface marker CD34. Therefore, the new chemokine-like proteins can be 15 used as diagnostic agents to identify CD34* progenitor cells in a sample population. For example, a solid substrate or matrix coated with a new chemokine-like protein can be used to separate out cells that are responsive to the protein from a sample of cells removed from a patient. After culturing or expansion of these CD34* cells ex vivo, these cells can be re-introduced into the patient following transplant, chemotherapy, or radiation therapy.

Furthermore, the new chemokine-like proteins can
25 be used as diagnostic screening agents to identify
patients with hyperproliferative diseases who would be
responsive to therapy with the new chemokine-like
proteins. For example, an in <u>vitro</u> assay, e.g., as
described above, is used to test whether a particular new
30 chemokine-like protein suppresses the proliferation of
myeloid cells, e.g., leukemic cells, in a sample taken
from a patient. A positive result, e.g., an inhibition
of proliferation of 50 percent compared to a control,

indicates that the patient can be treated using the new protein as a therapeutic agent to suppress the proliferation of myeloid leukemias.

Therapeutic Uses

The new chemokine-like proteins of the invention 5 can be administered to a patient as adjunctive agents before and/or during chemotherapy or radiation therapy to protect myeloid progenitor cells from the cytotoxic effects of the chemotherapeutic agents or radiation. 10 new proteins place myeloid cells into a myeloprotected, slow-cycling state, thereby inhibiting or decreasing cell damage that could otherwise be caused by cell-cycle active chemotherapy drugs such as cytosine arabinoside, 5-fluorouracil, or hydroxyurea. The use of the new 15 proteins also permits the administration of higher doses of chemotherapeutics without compromising the ability of the patient to generate mature functional blood cells. The new chemokines can be used to protect progenitor cells in either an in vivo or in vitro setting.

20 The new protein IL-8M3 can be administered to a patient to enhance progenitor cell proliferation by inhibition of the myelosuppressive activity of wild-type IL-8, or IL-8-based chemokines having myelosuppressive activity (e.g., IL-8M1 and IL-8M10). IL-8M3, or other chemokines that negatively regulate chemokine-mediated myelosuppression, can be administered to patients in whom myeloid cell proliferation is desirable, e.g., in patients having abnormally low numbers of cells derived from myeloid progenitors as a result of disease, e.g., leukemia or HIV infection, or as a side-effect of a therapeutic regimen such as chemotherapy.

The new chemokine-like proteins are administered to a patient in the same way as normal, wild-type chemokines are administered, e.g., they are injected

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intravenously or subcutaneously in a pharmaceutically acceptable carrier. For example, sterile saline or 30% acetonitrile/0.1% trifluoroacetic acid (ACN), as suggested for MIP-1¢ in Cooper et al. supra, can be used 5 as a carrier.

As described above, in <u>vivo</u> murine studies with IL-8M1 have shown that effective suppression of progenitor cell proliferation occurs at dosages of approximately 10.0 to 0.01 µg per animal, although 1.0 to 0.01 µg per animal is preferred. This translates into a dosage of approximately 0.5 to 500 µg/kg. Assuming the average human patient weighs 70 kgs, the effective amount for a human would be approximately 0.035 to 35.0 mg. Thus, a suitable dosage for therapy in a human patient is in the range of about 0.035 to 35.0 mg, with a preferred range of about 0.5 to 5 mg.

Note that in chemotherapy, specific protocols vary, and factors such as tumor size, growth rate, and location of the tumor all affect the course of adjunctive 20 therapy with the new proteins. Administration of chemotherapeutic agents, as well as the new chemokinelike proteins, requires knowledge of the extent of disease, the toxicity of previous treatment courses, and the timing of the expected drug toxicity.

The new chemokine-like proteins also can be used to inhibit hyperproliferative myeloid-based diseases such as chronic myelogenous leukemia, polycythemia vera, and hypermegakaryocytopoietic disorders. Hyperproliferative states in such disorders occur because the progenitor cells are unable to negatively regulate cell growth and replication. Administration of the new chemokine-like proteins is expected to inhibit cell replication resulting in the inhibition of the abnormal cell growth. This expected effect is based on indications that certain populations of myeloid leukemia cells are responsive to

the inhibitory effects of wild-type chemokines. Dosages of chemokine-like proteins for treating hyperproliferative myeloid based diseases would be similar to those dosages described above for use of the proteins as adjuncts to chemotherapy.

In addition, the new chemokine-like proteins can be used to prevent myeloid progenitor cells from becoming leukemic as the result of the administration of chemotherapeutic agents. The chemokine-like proteins are 10 administered in the same way described above.

The new chemokine-like proteins also can be used in a method of isolating progenitor cells for, for example, bone marrow transplantation. Bone marrow transplants require collection of a large number of

- 15 progenitor cells. Because progenitor cells are normally present in peripheral blood in very low numbers, isolation of a sufficient number of cells normally requires a bone marrow tap. The new chemokine-like proteins can be administered to the patient intravenously to mobilize progenitor cells from the bone marrow to the
- 20 to mobilize progenitor cells from the bone marrow to the peripheral blood, thus providing a sufficient number of progenitor cells in the peripheral blood and avoiding the need for a bone marrow tap. While intravenous administration of wild-type IL-8 also results in
- 25 progenitor cell mobilization, wild-type IL-8 also elicits neutrophil degranulation. The new chemokine-like proteins are advantageous for this use because they can elicit progenitor cell mobilization without eliciting neutrophil activation.

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Other Embodiments

It is to be understood that while the invention has been described in conjunction with the detailed description thereof, that the foregoing description is 5 intended to illustrate and not limit the scope of the invention, which is defined by the scope of the appended claims. Other aspects, advantages, and modifications are within the scope of the following claims.

WO 96/13587

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-51-

SEQUENCE LISTING

- (1) GENERAL INFORMATION:
 - (i) APPLICANT: Repligen Corporation
- (ii) TITLE OF INVENTION: Chemokine-Like Proteins and Methods of 5
 - (iii) NUMBER OF SEQUENCES: 70
- (iv) CORRESPONDENCE ADDRESS: (A) ADDRESSEE: Fish & Richardson P.C.
- (B) STREET: 225 Franklin Street 10
- (C) CITY: Boston
 - (D) STATE: MA
 - (E) COUNTRY: U.S.A. (F) ZIP: 02110-2804
- (V) COMPUTER READABLE FORM: 15
 - (A) MEDIUM TYPE: Floppy disk
 - (B) COMPUTER: IBM PC compatible (C) OPERATING SYSTEM: PC-DOS MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.30B
- (vi) CURRENT APPLICATION DATA: 20
 - (A) APPLICATION NUMBER: PCT/US95
 - (B) FILING DATE: 25-OCT-1995
 - (C) CLASSIFICATION:
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 - (A) APPLICATION NUMBER: US 08/482,111 (B) FILING DATE: 07-JUN-1995
 - (C) CLASSIFICATION:

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- 35 (2) INFORMATION FOR SEQ ID NO:1:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 72 amino acids
 - (B) TYPE: amino acid
- (C) STRANDEDNESS: single (D) TOPOLOGY: linear 40
 - (ii) MOLECULE TYPE: peptide
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:
 - Ser Ala Lys Glu Leu Arg Cys Cln Cys Ile Lys Thr Tyr Ser Lys Pro 1 10 15
- Phe His Pro Lys Phe Ile Lys Glu Leu Arg Val Ile Glu Ser Gly Pro 45

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His Cys Ala Asn Thr Glu Ile Ile Val Lys Leu Ser Asp Gly Arg Glu 45

Leu Cys Leu Asp Pro Lys Glu Asn Trp Val Gln Arg Val Val Glu Lys 50 55 Phe Leu Lys Arg Ala Glu Asn Ser 65 70

- (2) INFORMATION FOR SEQ ID NO:2:
- (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 73 amino acids
 - (B) TYPE: amino acid (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2: 15

Ala Ser Val Ala Thr Glu Leu Arg Cys Gln Cys Leu Gln Thr Leu Gln 1 10 15 Gly Ile His Pro Lys Asn Ile Gln Ser Val Asn Val Lys Ser Pro Gly 20 30

Pro His Cys Ala Gln Thr Glu Val Ile Ala Thr Leu Lys Asn Gly Arg 35 40 45 20 Lys Ala Cys Leu Asn Pro Ala Ser Pro Ile Val Lys Lys Ile Ile Glu 50 60

Lys Met Leu Asn Ser Asp Lys Ser Asn 65 25

- (2) INFORMATION FOR SEQ ID NO:3:
 - (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 73 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: single 30 (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:
- Ala Pro Leu Ala Thr Glu Leu Arg Cys Gln Cys Leu Gln Thr Leu Gln 1 15 35
 - Gly Ile His Leu Lys Asn Ile Gln Ser Val Asn Val Lys Ser Pro Gly 25 30
 - Pro His Cys Ala Gln Thr Glu Val Ile Ala Thr Leu Lys Asn Gly Gln 35 40 45
- Lys Ala Cys Leu Ann Pro Ala Ser Pro Het Val Lys Lys Ile Ile Glu 5040

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Lys Met Glu Lys Asn Gly Lys Ser Asn 65 70

- (2) INFORMATION FOR SEQ ID NO:4:
 - (1) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 73 amino acids (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4: 10
 - Ala Ser Val Val Thr Glu Leu Arg Cys Gln Cys Leu Gln Thr Leu Gln 1 5 10 15
 - Gly Ile His Leu Lys Asn Ile Gln Ser Val Asn Val Arg Ser Pro Gly 20 25 30
- Pro His Cys Ala Gln Thr Glu Val Ile Ala Thr Leu Lys Asn Gly Lys $35 \hspace{1cm} 40 \hspace{1cm} 45$ 15 Lys Ala Cys Leu Asn Pro Ala Ser Pro Met Val Gln Lys Ile Ile Glu 50 60

Lys Ile Leu Asn Lys Gly Ser Thr Asn 65

- (2) INFORMATION FOR SEQ ID NO:5:
 - (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 70 amino acids
 - (B) TYPE: amino acid
- (C) STRANDEDNESS: single (D) TOPOLOGY: linear 25
 - (ii) MOLECULE TYPE: peptide
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:
- Ala Glu Leu Arg Cys His Cys Ile Lys Thr Thr Ser Gly Ile His Pro 1 10 15 30

Lys Asn Ile Gln Ser Leu Glu Val Ile Gly Lys Gly Thr His Cys Asn 20 25 30

- Gln Val Glu Val Ile Ala Thr Leu Lys Asp Gly Arg Lys Ile Cys Leu 35
- Amp Pro Amp Ala Pro Arg Ile Lys Lys Ile Val Gin Lys Lys Leu Ala 50 60 35
 - Gly Asp Glu Ser Ala Asp
 - (2) INFORMATION FOR SEQ ID NO:6:
- (i) SEQUENCE CHARACTERISTICS: 40 (A) LENGTH: 70 amino acids
 (B) TYPE: amino acid

 - (C) STRANDEDNESS: single

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- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:
- Glu Ala Glu Glu Asp Gly Asp Leu Gln Cys Leu Cys Val Lys Thr Thr

Ser Cln Val Arg Pro Arg His Ile Thr Ser Leu Glu Val Ile Lys Ala 20 30

Gly Pro His Cys Pro Thr Ala Gln Leu Ile Ala Thr Leu Lys Asn Gly 35 40

Arg Lys Ile Cys Leu Asp Leu Gln Ala Pro Leu Tyr Lys Lys Ile Ile 50 50 10 Lys Lys Leu Leu Glu Ser 65 70

- (2) INFORMATION FOR SEQ ID NO:7:
- (i) SEQUENCE CHARACTERISTICS: 15 (A) LENGTH: 77 amino acids

 - (B) TYPE: amino acid (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide 20
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

Val Pro Leu Ser Arg Thr Val Arg Cys Thr Cys Ile Ser Ile Ser Aen 1 5 5 10 10 15

Gln Pro Val Asn Pro Arg Ser Leu Glu Lys Leu Glu Ile Ile Pro Ala 20 $$30\,$

Ser Gln Phe Cys Pro Arg Val Glu Ile Ile Ala Thr Met Lys Lys Lys 45 45

Gly Glu Lys Arg Cys Leu Asn Pro Glu Ser Lys Ala Ile Lys Asn Leu 50

Leu Lys Ala Val Ser Lys Glu Met Ser Lys Arg Ser Pro 65 75 30

- (2) INFORMATION FOR SEQ ID NO:8:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 78 amino acids
 - (B) TYPE: amino acid (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:
- Ala Gly Pro Ala Ala Ala Val Leu Arg Glu Leu Arg Cys Val Cys Leu 1 10 15 40

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Gln Thr Thr Gln Gly Val His Pro Lys Met Ile Ser Asn Leu Gln Val 20 25 30Phe Ala Ile Gly Pro Gln Cys Ser Lys Val Glu Val Val Ala Ser Leu 35 40 45

- Lys Asn Gly Lys Glu Ile Cys Leu Asp Pro Glu Ala Pro Phe Leu Lys 50 60 Lys Val Ile Gln Lys Ile Leu Asp Gly Gly Asn Lys Glu Asn 65 75
 - (2) INFORMATION FOR SEQ ID NO:9:
- 10 (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 81 amino acids
 - (B) TYPE: amino acid (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide 15
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

Gly Lys Glu Glu Ser Leu Asp Ser Asp Leu Tyr Ala Glu Leu Arg Cys

Met Cys Ile Lys Thr Thr Ser Gly Ile His Pro Lys Asn Ile Gln Ser 20 Leu Glu Val Ile Gly Lys Gly Thr His Cys Asn Gln Val Glu Val Ile

Ala Thr Leu Lys Asp Gly Arg Lys Ile Cys Leu Asp Pro Asp Ala Pro

Arg Ile Lys Lys Ile Val Gln Lys Lys Leu Ala Gly Asp Glu Ser Ala 65 70 75 80 25

(2) INFORMATION FOR SEQ ID NO:10:

(i) SEQUENCE CHARACTERISTICS: 30

Asp

- (A) LENGTH: 84 amino acids
 - (B) TYPE: amino acid

 - (C) STRANDEDNESS: single (D) TOPOLOGY: linear
- 35 (ii) MOLECULE TYPE: peptide
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

Asn Leu Ala Lys Gly Lys Glu Glu Ser Leu Asp Ser Asp Leu Tyr Ala 1 10 15

Glu Leu Arg Cys Met Cys Ile Lys Thr Thr Ser Gly Ile His Pro Lys Asn Ile Gln Ser Leu Glu Val Ile Gly Lys Gly Thr His Cys Asn Cln 35 40

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Val Glu Val Ile Ala Thr Leu Lys Asp Gly Arg Lys Ile Cys Leu Asp. 50 60

Pro Asp Ala Pro Arg Ile Lys Lys Ile Val Gln Lys Lys Leu Ala Gly 65 70

- Asp Glu Ser Ala 5
 - (2) INFORMATION FOR SEQ ID NO:11:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 94 amino acids
- (B) TYPE: amino acid 10 (C) STRANDEDNESS: Bingle
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide

 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:
- Ser Ser Thr Lys Gly Gln Thr Lys Arg Asn Leu Ala Lys Gly Lys Glu 1 10 15 15
 - Glu Ser Leu Amp Ser Amp Leu Tyr Ala Glu Leu Arg Cym Het Cym Ile 20 30
 - Lys Thr Thr Ser Cly Ile His Pro Lys Asn Ile Gln Ser Leu Glu Val \$45\$Ile Gly Lys Gly Thr His Cys Asn Gln Val Glu Val Ile Ala Thr Leu 50 $\,$

 - Lys hap Gly Arg Lys Ile Cys Leu hap Pro Asp Ala Pro Arg Ile Lys 65 70 80
- Lys lie Val Gln Lys Lys Leu Ala Gly Asp Glu Ser Ala Asp 9025
 - (2) INFORMATION FOR SEQ ID NO:12:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 78 amino acids
- (B) TYPE: amino acid 30
 - (C) STRANDEDNESS: single (D) TOPOLOGY: linear

 - (ii) MOLECULE TYPE: peptide
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:
- His Met Gln Pro Asp Ala Ile Asn Ala Pro Val Thr Cys Cys Tyr Asn 1 10 15 35
 - Phe Thr Asn Arg Lys Ile Ser Val Gln Arg Leu Ala Ser Tyr Arg Arg 25 \$25\$
- Ile Thr Ser Ser Lys Cys Pro Lys Glu Ala Val Ile Phe Lys Thr Ile 40 $^{\rm 45}$ 40
 - Val Ala Lys Glu Ile Cys Ala Asp Pro Lys Gln Lys Trp Val Gln Asp 50 60

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Ser Met Asp His Leu Asp Lys Gln Thr Gln Thr Pro Lys Thr 65 70 75

- (2) INFORMATION FOR SEQ ID NO:13:
 - (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 72 amino acids

 - (B) TYPE: amino acid (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:13: 10

Phe Ser Ala Ser Leu Ala Ala Asp Thr Pro Thr Ala Cys Cys Phe Ser 1 15

Tyr Thr Ser Arg Gln Ile Pro Gln Asn Phe Ile Ala Asp Tyr Phe Glu 20 25

Thr Ser Ser Gln Cys Ser Lys Pro Gly Val Ile Phe Leu Thr Lys Arg 35 40 15

Ser Arg Gln Val Cys Ala Asp Pro Ser Glu Glu Trp Val Gln Lys Tyr 50 60

Val Ser Asp Leu Glu Leu Ser Ala 20

- (2) INFORMATION FOR SEQ ID NO:14:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 66 amino acids (B) TYPE: amino acid
- (C) STRANDEDNESS: single 25
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:
- Pro Tyr Ser Ser Asp Thr Thr Pro Cys Cys Phe Ala Tyr Ile Ala Arg 1 10 15 30

Cys Ser Asn Pro Ala Val Val Phe Val Thr Arg Lys Asn Arg Gln Val 35 40 45

Cys Ala Asn Pro Glu Lys Lys Trp Tyr Arg Glu Tyr Ile Asn Ser Leu 50 60 35

Glu Met

- (2) INFORMATION FOR SEQ ID NO:15:
- (i) SEQUENCE CHARACTERISTICS: 40 (A) LENGTH: 65 amino acids
 - (B) TYPE: amino acid (C) STRANDEDNESS: single

- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:
- Gly Ser Asp Pro Pro Thr Ala Cys Cys Phe Ser Tyr Thr Ala Arg Lys $1 ext{1}$ 5 Leu Pro Arg Asn Phe Val Val Asp Tyr Tyr Glu Thr Ser Ser Leu Cys 25 30

Ser Gln Pro Ala Val Val Phe Gln Thr Lys Arg Ser Lys Gln Val Cys 35 40

Ala Asp Pro Ser Glu Ser Trp Val Gin Glu Tyr Val Tyr Asp Leu Glu 50 60 10

Leu 65

- (2) INFORMATION FOR SEQ ID NO:16:
- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 72 amino acids 15
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide 20
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:
 - Ser Ala Lys Asp Leu Gln Cys Gln Cys Ile Lys Thr Tyr Ser Lys Pro $1 ext{1}$ $5 ext{1}$
- Phe His Pro Lys Phe Ile Lys Glu Leu Arg Val Ile Glu Ser Gly Pro 20 25 30 25
 - His Cys Ala Asn Thr Glu Ile Ile Val Lys Leu Ser Asp Gly Arg Glu 40 45
 - Leu Cys Leu Asp Pro Lys Glu Asn Trp Val Gln Arg Val Val Glu Lys 50 50
- Phe Leu Lys Arg Ala Glu Asn Ser 30
 - (2) INFORMATION FOR SEQ ID NO:17:
 - (1) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 72 amino acids
 (B) TYPE: amino acid
- 35 (C) STRANDEDNESS: Bingle (D) TOPOLOGY: linear

 - (ii) MOLECULE TYPE: peptide
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:
- Ser Ala Lys Glu Leu Arg Cys Gln Cys Ile Lys Thr Tyr Ser Lys Pro 1 15 40

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Phe His Pro Lys Phe Ile Lys Glu Tyr Arg Arg Ile Glu Ser Gly Pro $\overset{\cdot}{}$ 20 $^{\cdot}$ 25

His Cys Ala Asn Thr Glu Ile Ile Val Lys Leu Ser Asp Gly Arg Glu 45

5 Leu Cys Leu Asp Pro Lys Glu Asn Trp Val Gln Arg Val Val Glu Lys
50 60

Phe Leu Lys Arg Ala Glu Asn Ser

- (2) INFORMATION FOR SEQ ID NO:18:
- 10 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 72 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- 15 (ii) MOLECULE TYPE: peptide
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

Ser Ala Lys Glu Leu Arg $^{\circ}$ ys Gln Cys Ile Lys Thr Tyr Ser Lys Pro 10 15

Phe His Pro Lys Phe Ile Lys Leu Glu Arg Val Ile Glu Ser Gly Pro 20 25 30 His Cys Ala Asn Thr Glu Ile Ile Val Lys Leu Ser Asp Gly Arg Glu 45 45

Leu Cys Leu Asp Pro Lys Glu Asn Trp Val Gln Arg Val Val Glu Lys
50 55 60

25 Phe Leu Lys Arg Ala Glu Asn Ser 65 70

- (2) INFORMATION FOR SEQ ID NO:19:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 72 amino acids (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:
- 35 Ser Ala Lys Glu Leu Arg Cys Gln Cys Ile Lys Thr Tyr Ser Lys Pro
 1 10 15

 Phe His Pro Lys Phe Ile Lys Glu Leu Arg Ala Ile Glu Ser Gly Pro
 20 30
 - 20 Z5 30

 His Cys Ala Asn Thr Glu Ile Ile Val Lys Leu Ser Asp Gly Arg Glu
 45
 45
 - Leu Cys Leu Asp Pro Lys Glu Asn Trp Val Cln Arg Val Val Glu Lys 50 60

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Phe Leu Lys Arg Ala Glu Asn Ser

- (2) INFORMATION FOR SEQ ID NO:20:
- (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 68 amino acids 5
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:20: 10

Ser Ala Lys Glu Leu Arg Cys Gln Cys Ile Lys Thr Tyr Ser Lys Pro 1 10 15

Phe His Pro Lys Phe Ile Lys Glu Leu Arg Val Ile Glu Ser Gly Pro 20 30

His Cys Ala Asn Thr Glu Ile Ile Val Lys Leu Ser Asp Gly Arg Glu 4515 Leu Cys Leu Asp Leu Gln Ala Pro Leu Tyr Lys Lys Ile Ile Lys Lys 50

Leu Leu Clu Ser

- 20 (2) INFORMATION FOR SEQ ID NO:21:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 64 amino acids
- (B) TYPE: amino acid (C) STRANDEDNESS: Bingle 25 (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:
- Ser Ala Lys Glu Leu Arg Cys Gln Cys Ile Lys Thr Tyr Ser Lys Pro 1 15 30

Phe His Pro Lys Phe Ile Lys Glu Leu Arg Val Ile Glu Ser Gly Pro $20 \ \ 25$

His Cys Ala Asn Thr Glu Ile Ile Val Lys Leu Ser Asp Gly Arg Glu 45 45

- Leu Cys Leu Asp Pro Lys Glu Asn Trp Val Gln Arg Val Val Glu Lys 55 60 35
 - (2) INFORMATION FOR SEQ ID NO:22:
 - (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 67 amino acids
- (B) TYPE: amino acid 40
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide

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- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:
- Ser Ala Lys Glu Leu Arg Cys Gln Cys Val Lys Thr Thr Ser Gln Val 1 10 15
- Arg Pro Arg His 11e Thr Ser Leu Glu Val 11e Lys Ala Gly Pro His 20 25
 - Cys Pro Thr Ala Cln Leu Ile Ala Thr Leu Lys Asn Cly Arg Lys Ile 35 40 45
 - Cys Leu Asp Leu Gln Ala Pro Leu Tyr Lys Lys Ile Ile Lys Lys Leu 50 55
- 10 Leu Glu Ser 65

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- (2) INFORMATION FOR SEQ ID NO:23:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 70 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:
- Glu Ala Glu Glu Asp Cly Asp Leu Arg Cys Leu Cys Val Lys Thr Thr 20
 - Ser Gln Val Arg Pro Arg His Ile Thr Ser Leu Glu Val Ile Lys Ala 20 25
- Gly Pro His Cys Pro Thr Ala Gln Leu Ile Ala Thr Leu Lys Asn Gly 35 40 25
 - Arg Lys Ile Cys Leu Asp Leu Gln Ala Pro Leu Tyr Lys Lys Ile Ile 50 60Lys Lys Leu Leu Glu Ser
- 30 (2) INFORMATION FOR SEQ ID NO:24:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 70 amino acids
 - (B) TYPE: amino acid (C) STRANDEDNESS: Bingle
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:
 - Glu Ala Glu Glu Asp Gly Asp Leu Gln Cys Leu Cys Val Lys Thr Thr 1 10 15
- Ser Gln Val Arg Pro Arg His Ile Thr Ser Leu Glu Val Ile Lys Ala 20 30 40

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Gly Pro His Cys Pro Thr Ala Gln Leu Ile Ala Thr Leu Lys Asn Gly . 45 $^{\rm 40}$

Arg Lys Leu Cys Leu Asp Pro Lys Glu Asn Trp Val Lys Lys Ile Ile 50 60

- Lys Lys Leu Leu Glu Ser
 - (2) INFORMATION FOR SEQ ID NO:25:
 - (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 70 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:
- Glu Ala Glu Glu Asp Gly Asp Leu Gln Cys Leu Cys Val Lys Thr Thr 15
 - Ser Gln Val Arg Pro Arg His Ile Thr Ser Leu Glu Val Ile Lys Ala 20 25 30 Gly Pro His Cys Pro Thr Ala Gln Leu Ile Ala Thr Leu Lys Asn Gly 35 40 45
 - Arg Lys Ile Cys Leu Asp Pro Asp Ala Pro Arg Ile Lys Lys Ile Ile 50 $\,$

Lys Lys Leu Leu Glu Ser

- 25 (2) INFORMATION FOR SEQ ID NO:26:
 - (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 70 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear 30
 - (ii) MOLECULE TYPE: peptide (x1) SEQUENCE DESCRIPTION: SEQ ID NO:26:
 - Glu Ala Glu Glu Asp Gly Asp Leu Gln Cys Leu Cys Val Lys Thr Thr
- Ser Gln Val Arg Pro Arg His Ile Thr Ser Leu Glu Val Ile Lys Ala 20 2535
 - Gly Pro His Cys Pro Thr Ala Gln Leu Ile Ala Thr Leu Lys Asn Gly
 45
 45
- Arg Lys Ala Cys Leu Asn Pro Ala Ser Pro Ile Val Lys Lys Ile Ile 50 $^{50}\,$ Lys Lys Leu Leu Glu Ser

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- (2) INFORMATION FOR SEQ ID NO:27:
 - (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 70 amino acids
 - (B) TYPE: amino acid STRANDEDNESS: single
- 5 (D) TOPOLOGY: linear

 - (ii) MOLECULE TYPE: peptide
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:
- Glu Ala Glu Glu Asp Gly Asp Leu Gln Cys Leu Cys Val Lys Thr Thr 10 Ser Gln Val Arg Pro Arg His Ile Lys Glu Leu Arg Val Ile Glu Ala 20 25 30
 - Gly Pro His Cys Pro Thr Ala Gln Leu Ile Ala Thr Leu Lys Asn Gly 35 40 45
- Arg Lys Ile Cys Leu Asp Leu Gln Ala Pro Leu Tyr Lys Lys Ile Ile 50 $60\,$ 15 Lys Lys Leu Leu Glu Ser 65
 - (2) INFORMATION FOR SEQ ID NO:28:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 70 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide 25
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:
 - Glu Ala Glu Glu Asp Gly Asp Leu Gln Cys Leu Cys Val Lys Thr Thr
- Ser Gln Val Gln Pro Gln His Ile Thr Ser Leu Glu Val Ile Lys Ala 20 25 30 30
 - Gly Pro His Cys Pro Thr Ala Gln Leu Ile Ala Thr Leu Lys Asn Gly
 - Gln Lys Ile Cys Leu Asp Leu Gln Ala Pro Leu Tyr Lys Lys Ile Ile 50 60
- Lys Lys Leu Leu Glu Ser 65 70 35
 - (2) INFORMATION FOR SEQ ID NO:29:
 - (1) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 310 base pairs (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear

	(ii) HOLECULE TYPE: DNA	
	(x1) SEQUENCE DESCRIPTION: SEQ ID NO:29:	0
	PROCESS COGGGTTCTG CGCCTCACGC CCCGGGCIGC IGII	
	CONTROL TEGTOGCCTT CGCCAGCGCT GAAGCTGAAG AAGATGGGGA 12	
5		
,	PARTICIPATE PARTICIPATE COCACTOCC CACTOCCAA CTGATAGCCA CGCTGAAGAA	•
	TOTAL COLD ATTIGCTING ACCINCAAGO COCCOTTAC AAGAAAATAA TTAAGAAACI SO	,,
	TTTGGAGAGT	10
	(2) INFORMATION FOR SEQ ID NO:30:	
10	(1) SEQUENCE CHARACTERISTICS: (A) LENGTH: 57 base pairs (B) TYPE: nucleic actid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
15	(ii) MOLECULE TYPE: DNA	
	(x1) SEQUENCE DESCRIPTION: SEQ ID NO:30:	57
	GRAGOTGRAG RAGATGGGGA COTGCAGTGC CTGTGTGTGA AGACCACCTC CORNOTO	•
	(2) INFORMATION FOR SEQ ID NO:31:	
20	(1) SEQUENCE CHARACTERISTICS: (A) LENGTH: 100 base pairs (B) TTPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
	(ii) HOLECULE TYPE: DNA	
25	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:31:	60
	COTOGOTATC AGTTGGGCAG TGGGGCAGTG GGGTCCGGCC TTGATCACCT CCAGGCTGGT	100
	GATGTGCCTG GGACGGACCT GGGAGGTGGT CTTCACACAC	
	(2) INFORMATION FOR SEQ ID NO:32:	
30	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 99 base pairs (B) TYPE nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: DNA	
3	5 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:32:	6
	5 (XI) SEQUENCE DEBOTATION OF GARGANIST AGGARANTIT GCTTGGACCT CTGCCCCATCTGCCCAACTGA TAGCCACGCT GAAGANISGA AGGARANTIT GCTTGGACCT	9

GCANGCCCCG CTGTACAAGA AAATAATTAA GAAACTTTT

	(2) INFORMATION FOR SEQ ID NO:33:	
5	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 40 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: DNA	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:33:	
	CTAACTCTCC AAAAGTTTCT TAATTATTTT CTTGTACAGC	40
LO	(2) INFORMATION FOR SEQ ID NO:34:	
15	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 207 base pairs (B) TYPE: nucleic scid (C) STRANDEDNESS: double (D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: DNA	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:34:	
	ATGACTGCTA AAGAACTTAG ATGTCAGTGC GTGAAGACCA CCTCCCCCCC	60
	CACATCACCA GCCTGGAGGT GATCAAGGCC GGACCCCACT GCCCCACTGC TCAGCTGATA	120
20	GCCACGCTGA AGANTGGAAG GAAANTTTGC TTGGACCTGC AAGCCCCGGT GTACAAGAAA.	180
	ATANTTANGA ANCTITTGGA GAGTTAG	207
	(2) INFORMATION FOR SEQ ID NO:35:	
	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 57 base pairs	
25	(B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
25	(B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (ii) HOLECULE TYPE: DNA	
25	(B) TYPE: nucleic acid (C) STRANDENNESS: single (D) TOPOLOGY: linear (ii) MOLECULE TYPE: DNA (xi) SEQUENCE DESCRIPTION: SEQ ID NO:35:	-
25 30	(B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (ii) MOLECULE TYPE: DNA (xi) SEQUENCE DESCRIPTION: SEQ ID No:35: ATGAGTGCTA AAGAACTTAG ATGTCAGTGC GTGRAGACCA CCTCCCAAGGT CCGTCCC	57
	(B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (ii) MOLECULE TYPE: DNA (xi) SEQUENCE DESCRIPTION: SEQ ID No:35: ATGAGTGCTA AAGAACTTAG ATGTCAGTGC GTGAAGACCA CCTCCCAAGT CCGTCCC (2) INFORMATION FOR SEQ ID NO:36:	57
	(B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (ii) MOLECULE TYPE: DNA (xi) SEQUENCE DESCRIPTION: SEQ ID No:35: ATGAGTGCTA AAGAACTTAG ATGTCAGTGC GTGRAGACCA CCTCCCAAGGT CCGTCCC	57
30	(B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (ii) HOLECULE TYPE: DNA (xi) SEQUENCE DESCRIPTION: SEQ ID No:35: ATGAGTGCTA AAGAACTTAG ATGTCAGTGC GTGAAGACCA CCTCCCAGGT CCGTCCC (2) INFORMATION FOR SEQ ID NO:36: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 219 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double	57
30	(B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (ii) MOLECULE TYPE: DNA (xi) SEQUENCE DESCRIPTION: SEQ ID NO:35: ATGAGTGCTA AAGAACTTAG ATGTCAGTGC GTGAAGACCA CCTCCCAGGT CCGTCCC (2) INFORMATION FOR SEQ ID NO:36: (1) SEQUENCE CHARACTERISTICS: (A) LENGTH: 219 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: linear (ii) MOLECULE TYPE: DNA (xi) SEQUENCE DESCRIPTION: SEQ ID NO:36:	
30	(B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (ii) MOLECULE TYPE: DNA (xi) SEQUENCE DESCRIPTION: SEQ ID NO:35: ATGAGTGCTA AAGAACTTAG ATGTCAGTGC GTGAAGACCA CCTCCCAGGT CCGTCCC (2) INFORMATION FOR SEQ ID NO:36: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 219 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: linear (ii) MOLECULE TYPE: DNA (xi) SEQUENCE DESCRIPTION: SEQ ID NO:36:	60

	ATTGTARAGC TTTCTGATGG AAGAGAGCTC TGTCTGGACC CCAAGGARAA CTGGGTGCAG	٠.
	AGGGTTGTGG AGAAGTTTTT GAAGAGGGCT GAGAATTCA 2	9
	(2) INFORMATION FOR SEQ ID NO:37:	
5	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 59 base pairs (B) TYPE: nucleic acid (C) STRANDENRESS: single (D) TOPLOOF: linear	
	(11) MOLECULE TYPE: DNA	
10	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:37:	
	ATGAGTGCTA AAGAACTTAG ATGTCAGTGC ATAAAGACAT ACTCCAAACC TTTCCACCC	59
	(2) INFORMATION FOR SEQ ID NO:38:	
15	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 99 base pairs (B) TYPE: nucleic eid (C) STRANDEDMESS: single (D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: DNA	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:38:	
20	CCATCAGAAA GCTTTACAAT AATTTCTGTG TTGGCGCAGT GTGGTCCACT CTCAATCACT	60
	CTCAGTTCTT TGATAAATTT GGGGTGGAAA GGTTTGGAG	99
	(2) INFORMATION FOR SEQ ID NO:39:	
25	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 99 base pairs (B) TTF2: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: DNA	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:39:	60
30		99
	AACTGGGTGC AGAGGGTTGT GGAGAAGTTT TTGAAGAGG	,,
	(2) INFORMATION FOR SEQ ID NO:40:	
3!	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 37 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
	(11) MOLECULE TYPE: DNA	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:40:	37
4	O TGAATTCTCA GCCCTCTTCA AAAACTTCTC CACAACC	٠,

	(2) INFORMATION FOR SEQ ID NO:41:	
5	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 219 base pairs (B) TYPE: nucleic acid (C) STRANDENESS: double (D) TOPOLOGY: linear	
	*	
	(ii) MOLECULE TYPE: DNA	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:41:	٤0
	ATGAGTGCTA AAGACCTGCA GTGTCAGTGC ATAAAGACAT ACTCCAAACC TTTCCACCCC	
10	ANATTTATCA ANGARCTGAG AGTGATTGAG AGTGGACCAC ACTGCGCCAA CACAGARATT I	20
	ATTGTAAAGC TAAGCGATGG AAGAGAGCTG TGTCTGGACC CCAAGGAAAA CTGGGTGCAG	180
	AGGGTTGTGG AGAAGTTTTT GAAGAGGGCT GAGAATTCA	219
	(2) INFORMATION FOR SEQ ID NO: 42:	
15	(1) SEQUENCE CHARACTERISTICS: (A) LENGTH: 47 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPLOGY: linear	
	(ii) MOLECULE TYPE: DNA	
20	(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 42:	47
	ATGAGTGCTA AAGACCTGCA GTGTCAGTGC ATAAAGACAT ACTCCAA	• /
	(2) INFORMATION FOR SEQ ID NO:43:	
25	(1) SEQUENCE CHARACTERISTICS: (A) LENGTH: 9 amino acids (B) TTPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: peptide	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:43:	
30	Ile Ala Thr Leu Lys Asn Gly Gln Lys 1	
	(2) INFORMATION FOR SEQ ID NO:44:	
35	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 11 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	

(ii) MOLECULE TYPE: peptide

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(xi) SEQUENCE DESCRIPTION: SEQ ID NO:44:

Ala Cys Leu Asn Pro Ala Ser Pro Ile Val Lys
1 5 10

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- (2) INFORMATION FOR SEQ ID NO:45:
 - (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 10 amino acids
 - (B) TYPE: amino acid
- STRANDEDNESS: single (C) STRANDEDNILL (D) TOPOLOGY: linear

 - (ii) MOLECULE TYPE: peptide
 - (x1) SEQUENCE DESCRIPTION: SEQ ID NO:45:
- Met Ser Ala Lys Glu Leu Arg Cys Gln Cys 10
 - (2) INFORMATION FOR SEQ ID NO:46:
 - (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 12 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 46:
 - Glu Ala Glu Glu Asp Gly Asp Leu Gln Cys Leu Gln
 - (2) INFORMATION FOR SEQ ID NO: 47:
 - (1) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 72 amino acids
- (B) TYPE: amino acid (C) STRANDEDNESS: not relevant 25
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:47:
- Ser Ala Lys Glu Leu Gln Cys Gln Cys Ile Lys Thr Tyr Ser Lys Pro 30
 - Phe His Pro Lys Phe Ile Lys Glu Leu Arg Val Ile Glu Ser Gly Pro 20 25
 - His Cys Ala Asn Thr Glu Ile Ile Val Lys Leu Ser Asp Gly Arg Glu 35 40
- Leu Cys Leu Amp Pro Lys Glu Amn Trp Val Gln Arg Val Val Glu Lys 50 60 35
 - Phe Leu Lys Arg Ala Glu Asn Ser 65
 - (2) INFORMATION FOR SEQ ID NO:48:
- (i) SEQUENCE CHARACTERISTICS: 40
 - (A) LENGTH: 52 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: not relevant

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- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:48:
- Ser Ala Lys Asp Leu Arg Cys Gln Cys Ile Lys Thr Tyr Ser Lys Pro 1 10 15

His Cys Ala Asn Thr Glu Ile Arg Val Val Glu Lys Phe Leu Lys Arg

Ala Glu Asn Ser 10

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- (2) INFORMATION FOR SEQ ID NO:49:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 72 amino acids (B) TYPE: amino acid
 - (C) STRANDEDNESS: not relevant
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:49:
- Ser Ala Lys Asp Leu Asn Cys Gln Cys Ile Lys Thr Tyr Ser Lys Pro 20

Phe His Pro Lys Phe Ile Lys Glu Leu Arg Val Ile Glu Ser Gly Pro 20 25

25 Leu Cys Leu Asp Pro Lys Glu Aen Trp Val Gln Arg Val Val Glu Lys 50 60

Phe Leu Lys Arg Ala Glu Asn Ser 65 70

- 30 (2) INFORMATION FOR SEQ ID NO:50:
 - (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 229 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA
 - (x1) SEQUENCE DESCRIPTION: SEQ ID NO:50:

CAAGCTTCAT ATGAGTGCTA AAGACCTGAA CTGTCAGTGC ATAAAGACAT ACTCCAAACC 60 TTTCCACCCC ARATTTATCA ARGANCTGAG AGTGATTGAG AGTGGACCAC ACTGCGCCAA 120

40 CACAGAAATT ATTGTAAAGC TAAGCGATGG AAGAGAGCTG TGTCTGGACC CCAAGGAAAA 180

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	CTCGGTGCAG AGGGTTGTGG AGARGTTTTT GAAGAGGGCT GAGAATTCA	229
	(2) INFORMATION FOR SEQ ID NO:51:	
5	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 37 base pairs (B) TYPE: nucleic seid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: protein	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:51:	37
10	CANGCTTCAT ATGAGTGCTA AAGACCTGAA CTGTCAG	31
	(2) INFORMATION FOR SEQ ID NO:52:	
15	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 228 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
	(ii) HOLECULE TYPE: DNA	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:52:	60
	CARGCTCATA TGAGTGCTAA AGAACTGCAG TGTCAGTGCA TAAAGACATA CTCCAAACCT	- 60
20	TTCCACCCCA AATTTATCAA AGAACTGAGA GTGATTGAGA GTGGACCACA CTGCGCCCAAC	120
	ACAGAAATTA TTGTAAAGCT AAGCGATGGA AGAGAGCTGT GTCTGGACCC CAAGGAAAAC	228
	TGGGTGCAGA GGGTTGTGGA GAAGTTTTTG AAGAGGGCTG AGAATTCA	220
	(2) INFORMATION FOR SEQ ID NO:53:	
25	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 40 base pairs (B) TYPE: nucleic acid (C) STRANDENNES: single (D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: DNA	
30	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:53:	40
	CAAGCTCATA TGAGTGCTAA AGAACTGCAG TGTCAGTGCA	
	(2) INFORMATION FOR SEQ ID NO:54:	
35	(i) SEQUENCE CHARACTERISTICS: (A) LEGISTIC 228 base pairs (B) TYP: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
	ALLA MOLECULE TYPE: DNA	

(x1) SEQUENCE DESCRIPTION: SEQ ID NO:54:

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	CAAGCTTCAT ATGAGTGCTA RAGATCTGCG TTGTCAGTGC ATRAAGACAT ACTCCAAACC	60	
	TITCCACCCC AAATTTATCA AAGAACTGAG AGTGATTGAG AGTGGACCAC ACTGCGCCAA 1	20	
	CACAGAAATT ATTGTAAAGC TAAGCGATGG AAGAGAGCTG TGTCTGGACC CCAAGGAAAA 1	.80	
	CTGGCTGCAG AGGGTTGTGG AGAGTTTTT GAAGAGGGCT GAGAATTCA	228	
5	(2) INFORMATION FOR SEQ ID NO:55:		
10	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 37 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPCLOGY: linear		
	(ii) MOLECULE TYPE: DNA		
	(x1) SEQUENCE DESCRIPTION: SEQ ID NO:55:		
	CAAGCTTCAT ATGAGTGCTA AAGATCTGCG TTGTCAG	37	
	(2) INFORMATION FOR SEQ ID NO:56:		
15	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 231 base pairs (B) TYPE: nucleic acid (C) STRANDEDIRESS: single (D) TOPOLOGY: linear		
20	(ii) MOLECULE TYPE: DNA		
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:56:	60	
	ATGAGTGCTA AAGAACTTAG ATGTCAGTGC ATAAAGACAT ACTCCAAACC TTTCCACCCC		
	ARATTTATCA ARGRATACAG ACGTATTGAG AGTGGACCAC ACTGCGCCAA CACAGARATT	120	
	ATTGTARAGC TARGCGATGG ANGAGAGCTG TGTCTGGACC CCARGGARAR CTGGGTGCAG	231	
25	AGGGTTGTGG AGAAGTTTTT GAAGAGGGCT GAGAATTCAT AGTAAGGATC C	231	
	(2) INFORMATION FOR SEQ ID NO:57:		
30	(1) SEQUENCE CHARACTERISTICS: (A) LENGTH: 313 base pairs (B) TYPE: nucleic acid (C) STRANDENNESS: single (D) TOPOLOGY: linear		
	(ii) MOLECULE TYPE: DNA		
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:57:	60	
	CCCCAGCATG AGCTCCGCAG CCGGGTTCTG CGCCTCACGC CCCGGGCTGC TGTTCCTCGG	120	
35	GTTGCTGCTC CTGCCACTTG TGGTCGCCTT CGCCAGCGCT GAAGCTGAAG AAGATGGGGA	195	,
	CCTGCAGTGC CTGTGTGTGA AGACCACCTC CCAGGTCCGT CCCAGGCACA TCACCAGCCT	190	,
	GGAGGTGATC AAGGCCGGAC CCCACTGCCC CACTGCCCAA CTGATAGCCA CGCTCAAGAA	240	٠.

TGGAAGGAAA GCTTGCTTGA ACCCGGCATC CCCGATTGTC AAGAAAATAA TTAAGAAACT 300

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	TTTGCAGAGT TAG	313.
	(2) INFORMATION FOR SEQ ID NO:58:	
5	(i) SEQUENCE CHARACTERISTICS: (A) LENOTH: 83 base pairs (B) TTPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
	(11) MOLECULE TYPE: DNA	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:58:	60
10	CTAACTCTCC ARAAGTTTCT TAATTATTTT CTTGACAATC GGGGATGCCG GGTTCAAGCA	83
	AGCTTTCCTT CCATTCTTGA GCG	63
	(2) INFORMATION FOR SEQ ID NO:59:	
15	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 234 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: DNA	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:59:	
20	GTGGATATCA AGCTTCATAT GGAAGCTGAA GAAGATGGGG ACCTGCAGTG CCTGTGTGTG	60
	AAGACCACCT CCCAGGTCCA GCCCCAGCAC ATCACCAGCC TGGAGGTGAT CAAGGCCGG	120
	CCCCACTGCC CCACTGCCCA ACTGATAGCC ACGCTGAAGA ATGGACAGAA AATTTGCTTC	234
	GACCTGCANG CCCCGCTGTA CANGANANTA ATTANGANAC TTTTGCAGAG TTAG	234
	(2) INFORMATION FOR SEQ ID NO:60:	
25	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 67 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
30		
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:60:	c 60
	TEGRAGETER AGRAGATEGE GACCTECAGT GCCTGTGTGT GARGACCACC TCCCAGGTC	67
	AGCCCCA	
	(2) INFORMATION FOR SEQ ID NO:61:	
3	5 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 75 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	

40 (ii) MOLECULE TYPE: DNA

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	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:61:	
	GGCAGTGGGG CAGTGGGGTC CGGCCTTGAT CACCTCCAGG CTGGTGATGT GCTGGGGCTG	60
		75
	(2) INFORMATION FOR SEQ ID NO:62:	
_	(1) SEQUENCE CHARACTERISTICS:	
5	(A) LENGTH: 88 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
10	(11) MOLECULE TYPE: DNA	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:62:	
	GGCCGGACCC CACTGCCCCA CTGCCCAACT GATAGCCACG CTGAAGAATG GACAGAAAAT	60
	TTGCTTGGAC CTGCAAGCCC CGCTGTAC	88
	(2) INFORMATION FOR SEQ ID NO:63:	
15	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 73 amino acids (B) TYPE: amino acid (C) STRANDEDURSS: not relevant (D) TOPOLOGY: linear	
20	(ii) MOLECULE TYPE: protein	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:63:	
	Met Ser Ala Lys Glu Leu Gln Cys Gln Cys Ile Lys Thr Tyr Ser I	ys
	1 5	
25	Pro Phe His Pro Lys Phe Ile Lys Glu Leu Arg Val Ile Glu Ser C 20 25 30	,
	Pro His Cys Ala Asn Thr Glu Ile Ile Val Lys Leu Ser Asp Gly 1 35 40	
	Glu Leu Cys Leu Asp Pro Lys Glu Asn Trp Val Gln Arg Val Val (50 60	Glu
30	Lys Phe Leu Lys Arg Ala Glu Asn Ser 65 70	
	(2) INFORMATION FOR SEQ ID NO:64:	
35	(1) SEQUENCE CHARACTERISTICS: (A) LENGTH: 53 maino acids (B) TYPE: amino acid (C) STRANDEDNESS: not relevant (D) TOPOLOGY: linear	

(ii) MOLECULE TYPE: protein

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(xi) SEQUENCE DESCRIPTION: SEQ ID NO:64:

Met Ser Ala Lys Asp Leu Arg Cys Gln Cys Ile Lys Thr Tyr Ser Lys

Pro Phe His Pro Lys Phe Ile Lys Glu Leu Arg Val Ile Glu Ser Gly 20 25 30 5

Pro His Cys Ala Asn Thr Glu Ile Arg Val Val Glu Lys Phe Leu Lys 35 40 45

Arg Ala Glu Asn Ser

- 10 (2) INFORMATION FOR SEQ ID NO:65:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 73 amino acids
 - (B) TYPE: amino acid (C) STRANDEDNESS: not relevant
- (D) TOPOLOGY: linear 15
 - (ii) MOLECULE TYPE: protein
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:65:

Met Ser Ala Lys Asp Leu Asn Cys Gln Cys Ile Lys Thr Tyr Ser Lys 1 5 10

Pro Phe His Pro Lys Phe Ile Lys Glu Leu Arg Val Ile Glu Ser Gly 20 25 30 20

Pro His Cys Ala Asn Thr Glu Ile Ile Val Lys Leu Ser Asp Gly Arg 35 40 45

Glu Leu Cys Leu Asp Pro Lys Glu Asn Trp Val Gln Arg Val Val Glu 50 55 60

Lys Phe Leu Lys Arg Ala Glu Asn Ser 65 70

- (2) INFORMATION FOR SEQ ID NO:66:
 - (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 31 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:66: 35
 - Met Ser Ser Ala Ala Gly Phe Cys Ala Ser Arg Pro Gly Leu Leu Phe 1 5 10 15

Leu Cly Leu Leu Leu Pro Leu Val Val Ala Phe Ala Ser Ala 20 25 30

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- (2) INFORMATION FOR SEQ ID NO: 67:
 - (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 68 amino acids
 - (B) TYPE: amino acid
- - STRANDEDNESS: single (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (x1) SEQUENCE DESCRIPTION: SEQ ID NO: 67:
- Met Ser Ala Lys Glu Leu Arg Cys Gln Cys Val Lys Thr Thr Ser Gln
- 10 Val Arg Pro Arg His Ile Thr Ser Leu Glu Val Ile Lys Ala Gly Pro 20 25
 - His Cys Pro Thr Ala Gln Leu Ile Ala Thr Leu Lys Asn Gly Arg Lys 45 45
- Ile Cys Leu Asp Leu Cln Ala Pro Leu Tyr Lys Lys Ile Ile Lys Lys 50 55 15
 - Leu Leu Glu Ser 65
 - (2) INFORMATION FOR SEQ ID NO:68:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 71 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide 25
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:68:
 - Met Glu Ala Glu Glu Asp Gly Asp Leu Gln Cys Leu Cys Val Lys Thr 1 5 10
 - Thr Ser Gln Val Arg Pro Arg His Ile Thr Ser Leu Glu Val Ile Lys 20 25
 - Ala Cly Pro His Cys Pro Thr Ala Gln Leu Ile Ala Thr Leu Lys Asn 35 40
 - Gly Arg Lys Ile Cys Leu Asp Leu Gln Ala Pro Leu Tyr Lys Lys Ile 50 60
- lle Lys Lys Leu Leu Glu Ser 35
 - (2) INFORMATION FOR SEQ ID NO:69:
 - (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 73 amino acids (B) TYPE: amino acid 40
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide

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- (x1) SEQUENCE DESCRIPTION: SEQ ID NO:69:
- Met Ser Ala Lys Glu Leu Arg Cys Gln Cys Ile Lys Thr Tyr Ser Lys
- Pro Phe His Pro Lys Phe 11e Lys Glu Leu Arg Val Ile Glu Ser Gly 20 25
 - Pro His Cys Ala Asn Thr Glu Ile Ile Val Lys Leu Ser Asp Gly Arg 35 40 45
 - Glu Leu Cys Leu Asp Pro Lys Glu Asn Trp Val Gln Arg Val Val Glu 50 60
- Lys Phe Leu Lys Arg Ala Glu Asn Ser 65 10
 - (2) INFORMATION FOR SEQ ID NO:70:
 - (i) SEQUENCE CHARACTERISTICS:

 - (A) LENGTH: 73 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:70:
- Met Ser Ala Lys Asp Leu Gln Cys Gln Cys Ile Lys Thr Tyr Ser Lys 20
 - Pro Phe His Pro Lys Phe Ile Lys Glu Leu Arg Val Ile Glu Ser Gly 20 25
 - Pro His Cys Ala Asn Thr Glu Ile Ile Val Lys Leu Ser Asp Gly Arg 35 40 45
 - Glu Leu Cys Leu Asp Pro Lys Glu Asn Trp Val Gln Arg Val Val Glu 50 60
 - Lys Phe Leu Lys Arg Ala Glu Asn Ser 65 70

CLAIMS

- A protein having the amino acid sequence of a wild-type chemokine having four cysteine residues, with the following modifications:
- a) amino acids DLQ as the 3 contiguous amino acids on the N-terminus side of and proximal to the firs: cysteine residue of the wild-type chemokine; and
- b) amino acids ELXVX₁X₂X₃X₄X₅X₆ as the 10 amino acids on the N-terminus side of and proximal to the third 10 cysteine of the wild-type chemokine, wherein X is any amino acid, and X₁X₂X₃X₄X₅X₆ can be any six consecutive amino acids on the N-terminus side of and proximal to the third cysteine of the wild-type chemokine.
- A protein of claim 1 having the amino acid
 sequence SAKDLQCQCIKTYSKPFHPKFIKELRVIESGPHCANTELIVKLS-DGRELCLDPKENWVQRVVEKFLKRAENS (SEQ ID NO:16).
 - 3. A protein having the amino acid sequence of a wild-type chemokine having four cysteine residues, with the following modifications:
- 20 a) amino acids DLN as the 3 contiguous amino acids on the N-terminus side of and proximal to the first cysteine residue of the wild-type chemokine; and
- b) amino acids ELXVX₁X₂X₃X₄X₅X₆ as the 10 amino acids on the N-terminus side of and proximal to the third cysteine of the wild-type chemokine, wherein X is any

amino acid, and $x_1x_2x_3x_4x_5x_6$ can be any six consecutive amino acids on the N-terminus side of and proximal to the third cysteine of the wild-type chemokine.

- A protein of claim 3 having the amino acid
 sequence SAKDLNCQCIKTYSKPFHPKFIKELRVIESGPHCANTEIIVKLS-DGRELCLDPKENWVQRVVEKFLKRAENS (SEQ ID NO: 49).
 - 5. A protein having the amino acid sequence of a wild-type chemokine having four cysteine residues, with the following modifications:
- a) amino acids ELQ as the 3 contiguous amino acids on the N-terminus side of and proximal to the first cysteine residue of the wild-type chemokine; and
- b) amino acids ELXVX₁X₂X₃X₄X₅X₆ as the 10 amino acids on the N-terminus side of and proximal to the third 15 cysteine of the wild-type chemokine, wherein X is any amino acid, and X₁X₂X₃X₄X₅X₆ can be any six consecutive amino acids on the N-terminus side of and proximal to the third cysteine of the wild-type chemokine.
- A protein of claim 5 having the amino acid
 sequence SAKELQCQCIKTYSKPFHPXFIKELRVIESGPHCANTEIIVKLS-DGRELCLDPKENWVQRVVEKFLKRAENS (SEQ ID NO: 47).

- 7. A protein having the amino acid sequence of a wild-type chemokine having four cysteine residues, with the following modifications:
- a) amino acids DLR as the 3 contiguous amino acids 5 on the N-terminus side of and proximal to the first cysteine residue of the wild-type chemokine; and
- b) amino acids ELXVX₁X₂X₃X₄X₅X₆ as the 10 amino acids on the N-terminus side of and proximal to the third cysteine of the wild-type chemokine, wherein X is any 10 amino acid, and X₁X₂X₃X₄X₅X₆ can be any six consecutive amino acids on the N-terminus side of and proximal to the third cysteine of the wild-type chemokine.
 - A protein of any one of claims 1, 3, 5, or 7,
 wherein amino acid X is R, N, E, or Q.
- 9. A protein of claim 7 having the amino acid sequence SAKDLRCQCIKTYSKPFHPKFIKELRVIESGPHCANTEIIVKLS-DGRELCLDPKENWVQRVVEKFLKRAENS (SEQ ID NO: 48).
- 10. A protein having the amino acid sequence of a wild-type chemokine having four cysteine residues, with 20 the following modifications:
 - a) amino acids ELR as the 3 contiguous amino acids on the N-terminus side of and proximal to the first cysteine residue of the wild-type chemokine; and

- b) amino acids XDLQ as the 4 amino acids on the Cterminus side of and proximal to the fourth cysteine of the wild-type chemokine; wherein X can be any amino acid.
- 11. A protein of claim 10, wherein amino acid \boldsymbol{x} 5 is L.
 - 12. A protein of claim 10 having the amino acid sequence SAKELRCQCVKTTSQVRPRHITSLEVIKAGPHCPTAQLIATLKN-GRKICLDLQAPLYKKIIKKLLES (SEQ ID NO:22).
- 13. A protein having the amino acid sequence of a 10 wild-type chemokine having four cysteine residues, with the following modifications:
 - a) amino acids DLQ as the 3 contiguous amino acids on the N-terminus side of and proximal to the first cysteine residue of the wild-type chemokine; and
- b) amino acids ACLNPASPIVK (SEQ ID NO:44) replacing 11 amino acids including the fourth cysteine of the wild-type chemokine, wherein the C in ACLNPASPIVK corresponds to the fourth cysteine residue.
- 14. A protein of claim 13 having the amino acid 20 sequence EAEEDGDLQCLCVKTTSQVRPRHITSLEVIKAGPHCPTAQLIATLKN-GRKACLNPASPIVKKIIKKLLES (SEQ ID NO:26).

- 15. A protein having the amino acid sequence of a wild-type chemokine having four cysteine residues, with the following modifications:
- a) amino acids DLQ as the 3 contiguous amino acids
 on the N-terminus side of and proximal to the first
 cysteine residue of the wild-type chemokine; and
- b) amino acids IATLKNGQK (SEQ ID NO:43) and Z as
 the 10 amino acids on the N-terminus side of and proximal
 to the fourth cysteine of the wild-type chemokine,
 10 wherein Z is any amino acid.
 - 16. A protein of claim 15 having the amino acid sequence EAEEDGDLQCLCVKTTSQVQPQHITSLEVIKAGPHCPTAQLIATLKNGQK ICLDLQAPLYKKIIKKLLES (SEQ ID NO:28).
- 17. A protein of any one of claims 1, 3, 5, 7,
 10, 13, or 15, wherein said wild-type chemokine is interleukin-8 (IL-8), melanoma-growth stimulating factor (GRO-α), macrophage inflammatory protein-2 alpha (MIP-2α), MIP-1α, platelet factor 4 (PF4), neutrophil
 20 activating peptide-2 (NAP-2), Neutrophil Activating protein (78 amino acids in length) (ENA-78), platelet basic protein (PBP), connective tissue-activating peptide III (CTAP-III), β-thromboglobulin (βTG), gamma interferon inducible protein 10 (γIP-10), macrophage chemotactic and

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activating factor (MCAF), or Regulated on Activation, Normal T-cell Expressed and Secreted protein (RANTES).

- 18. A protein of claim 1, wherein said wild-type chemokine is interleukin-8 (IL-8).
- 5 19. A protein of claim 10, wherein said wild-type chemokine is platelet factor 4 (PF4).
 - 20. The use of a protein of claim 1 for the manufacture of a medicament for suppressing proliferation of an actively dividing myeloid cell.
- 21. A protein of claim 1 for use as an adjunctive agent in chemotherapy or radiation therapy.
 - 22. A protein of claim 1 for use in treating a hyperproliferative myeloid disease.
- 23. The use of a protein of claim 1 for the 15 manufacture of a medicament for treating a hyperproliferative myeloid disease.
 - 24. A protein of claim 22 or 23, wherein said disease is chronic myelogenous leukemia, polycythemia vera, or a hypermegakaryocytopoietic disorder.

25. A method of detecting CD34* myeloid cells in a sample of cells comprising

obtaining a sample of cells,

contacting the sample with a protein of claim 1 5 under conditions that allow said protein to bind to any CD34⁺ myeloid cells in the sample to form bound complexes, and

detecting any bound complexes in the sample as an indication of the presence of CD34⁺ myeloid cells in the 10 sample.

26. A method of isolating CD34⁺ myeloid cells from a sample of cells comprising obtaining a sample of cells,

contacting the sample with a protein of claim 1

15 under conditions that allow said protein to bind to any

CD34⁺ myeloid cells in the sample to form bound

complexes,

removing any bound complexes from the sample, and separating CD34⁺ myeloid cells from said bound complexes 20 to isolate the CD34⁺ myeloid cells from the sample.

27. A nucleic acid having the sequence of SEQ ID NO:34, SEQ ID NO:41, SEQ ID NO:50, SEQ ID NO:52, SEQ ID NO:54, SEQ ID NO:56, SEQ ID NO:57, or SEQ ID NO:59.

- 28. A protein having the amino acid sequence SAKELRCQCIKTYSKPFHPKFIKEYRRIESGPHCANTEIIVKLSDGRELCLDPKENW VQRVVEKFL (SEQ ID NO:17).
- 29. A protein of claim 28 for use in enhancing 5 myeloid cell proliferation by inhibition of IL-8 mediated myelosuppression.
- 30. A protein having the amino acid sequence of a wild-type chemokine having four cysteine residues, the amino acid sequence comprising a heterologous active

 10 domain selected from the group consisting of: ELR, DLN, ELQ, DLR, ELRV, DLQ, IATLKNGQK (SEQ ID NO:43), and ACLNPASPIVK (SEQ ID NO:44).
- 31. A protein of claim 30, wherein the heterologous active domain replaces a number of amino 15 acids identical to the number of amino acids in the active domain on the N-terminus side of and proximal to the first cysteine residue of the wild-type chemokine.
- 32. A protein of claim 30, wherein the heterologous active domain replaces a number of amino 20 acids identical to the number of amino acids in the active domain on the N-terminus side of and within 11 amino acids of the third cysteine residue of the wild-type chemokine.

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- 33. A protein of claim 30, wherein the heterologous active domain replaces a number of amino acids identical to the number of amino acids in the active domain on the N-terminus side of and within 11 amino acids of the fourth cysteine residue of the wild-type chemokine.
- 34. A protein of claim 30, wherein the heterologous active domain replaces a number of amino acids identical to the number of amino acids in the active domain on the C-terminus side of and within 11 amino acids of the fourth cysteine residue of the wild-type chemokine.

SSTWOOTKRNLAKGKEBSLDSDLYAELRGWCIKTTS. GIHPKHIQSLEVIGKGTHGNQVEVIATL. KDGRKIGLDPDAPRIKKIVQKKLAGDESAD (SBQ ID NO:11) AP-III NLAKGKEESLDSDLYAELRCMCIKITS,GIHPWIQSLEVIGKGTHCRQVEVIATL.KOGRKICLDPDAPRIKKIVQKKLAGDESA (SEQ ID NO:10) HNOPDAINAPVTCCYNF.TNRKISVQRLASYRRITSSKCPR.EAVI..FKTIVAKEICADPKQKHVQDSM.DHLDKQTQTPKT (SEQ ID NO:12) GRESIDSDLYABIRGMCIKTTS.GIHPKNIQSLEVIGKGTHCNQVEVIATL.KDGRKICLDPDAPRIKKIVQKKLAGDESAD (SRQ ID NO:9) AGPAANURELRCVCLQTTQ.GVHPKMISNLQVFAIGPQGSKVEVVASL.KNGKEICLDPEAPFLKKVIGKILDGGNKBN (SBQ ID NO:8) PSASLAADITTACCFSY., ISRQIPQNF1ADYF. ETSSQCSKP.GVI..FLTKRSRQVCADPSEEWVQKYVSDLELSA (SBQ ID NO:13) VPLSRTYRCTCISISNOPVNPRSLEKLEIIPASGFCPRVEIIATMKKKGEKRCLNPESKAIKMLLKAVSKEMSKRSP (SEQ ID NO:7) ASVATELRCÇCLÇTLQ.GIHPKNIQSVNVKSPQPHCAQTEVIATI..KNGRKACLNPASPIVKKIIEKMLNSDKSN (SEQ ID NO:2) APLATELRCQCLQTLQ.GIHLKNIQSVNVKSFGPHCAQTEVIATL. KNCQKACLNPASPWVKXIIENMEKNGKSN (SEQ ID NO:3) ASVVTELRCQCLQTLQ.GIHLKNIQSVNVRSPQPHCAQTEVIATL.KNGKKACLNPASPMVQXIIEKILNKGSTN (SEQ ID NO:4) PYSSDTTPCCFAY.IARPLDRAHIKEYF.YTSGKGSNP.AVV..FVTRKNRQVCANPEKKWYREYINSLEM (SEQ ID NO:14) GSDPPTACCFSY.TARKLPRNFVVDYY.ETSSLCSQP.AVV..FQTKRSKQVCADPSESWVQEYVYDLEL (SEQ ID NO:15) .. 8 mono SAKBLACQCIKTYSKPPHPKFIKELKVIESGPHCANTEIIVKL. SDGRELCLDPKENWVQRVVEKPLKRAENS (SEQ ID NO:1) AELACHCIKTTS.GIHPKNIQSLEVIGKGTHCNQVEVIATL.KDGRKICLDPDAPRIKKIVQKKLAGDESAD (SEQ ID NO:5) EAREDGDLQCLCVKTTS.QVRPRHITSLEVIKAGPHCPTAQLIATL.KNGRKICLDLQAPLYKKIIKKLLES (SEQ 1D NO:6) P-1B MTES P-10 IA 78 .b-1 .P10 ٩ Ģ 1_{P2} ğ 6 5

Fig. 2

SAKELGCQCIKTYSKPPHPKFIKELRVIESGPHCNNTE11VKLSDGRELCLDPKENWYQRVVEKFLKRAENS (SEQ 1D NO:47) SNADAROGIKTYSKPPHPKFIKALAVIESGPHCANTEIIVKLSDGRELCLDPKENWORWUEKFLKRAENS (SEQ ID NO:48) SAK<u>DLN</u>CQCIKTYSKPFHPKFIKELRVIESGPHCANTEIIVKLSDGRELCLDPKENWQRVUEKFLKRAENS (SEQ 1D NO:49) EAEEDGDLGCLCVKTTSQVQPQHITSLEVIKAGPHCPTAGAIAATLKNGQKICLDLGAPLYKKIIKKLLES (SEQ ID NO:28) EAREDGDLQCLCVKTTSQVRPRHIKELRYLEAGPHCPTAQLIATLKNGRKICLDLQAPLYKKIIKKLLES (SEQ ID NO:27) SAKELRCQCIKTYSKPFHPKFIKLERVIESGPHCANTEIIVKLSDGRELCLDPKENWVQRVVEKFLKRAENS (SEQ ID NO:18) SAKELROQCIKTYSKPPHPKFIKELRELESGPHCANTEIIVKLSDGRELCLDPKENWYQRVVEKFLKRAENS (SEQ ID NO:19) EAEEDGDLQCLCVKTTSQVRPRHITSLEVIKAGPHCPTAQLIATLKNGRKICLDDDAERLKKIIKKLLES (SEQ ID NO:25) EAEEDGDLQCLCVKTTSQVRPRHITSLEVIKAGPHCV1A0.1ATLKNGRKA<u>CLARASFLVE</u>KIIKKLLES (SEQ 1D NO:26) EAREBOGDLĄCLCVKTTSQVRPRHITSLEVIKAGPHCPTAQ:IATLKNGKK<u>LGLDDKENINUK</u>KIIKKLLES (SEQ ID NO:24) SAKDLOCQCIKTYSKPPHPKFIKELRVIESGPHCANTEIIVKLSDGRELCLDPKENWYGRVVEKFLKRAENS (SEQ ID NO:16) (SEQ ID NO:17) EAEEDGDLECLCWKTTSQVRPRHITSLEVIKAGPHCPTAQLIATLKNGRKICLDLAAPLYKKIIKKLLES (SEQ ID NO:23) (SEQ ID NO:1) EAREDG**DLA**CLCVKTTSQVRPRHITSLEVIKAGPHCPTAQLIATLKNGRKICL**DLA**APLYKKIIKKLLES (SEQ ID NO:6) SAKBLRCOCUKTISQURPRHITSLEVIKAGPHCPTAQLIATLKNGRKICLDLGAPLYRKIIKKLLES (SEQ 1D NO:22) SAKELRCGCIKTYSKPFHPRFIKELRUIESGPHCANTEIIVKLSDGRELCLDPRENHVQRVVEKFLKRAENS sak**el**rcqciktyskpphpkfike<u>y</u>r<u>b</u>ibsgphcanteiivklsdgrelcldpkenwugruvekflkraens SAKELROQCIKTYSK PFH PKFIKELRVI ESGPHCANTEI I VKLSDGREL<u>CLDLGAPLYKKI IKKLLES</u> SAKELRCQCIKTYSKPFHPKFIKELRVIESOPHCANTEIIVKLSDGRELCLDPKENNVQRVVEK 426 413 414 421

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Fig. 3A (SEQ ID NO:29)

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Fig. 3B (SEQ ID NO:34)

	(SEQ ID NO:35)
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70	TACTCACGATTTCTTGAATCTACAGTCACGCACTTCTGGTGGAGGGTCCAGGCAGG
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	CACATCACCAGCCTGGAGGTGATCAAGGCCGGACCCCACTGCCCCACTGCTCAGCTGATA 19
131	GTGTAGTGGTCGGACCTCCACTAGTTCCGGCCTGGGGTGACGGGTTAC
	HITSLEVIKAGPHCPTAQLI -
	GCCACGCTGAAGAATGGAAGGAAAATTTGCTTGGACCTGCAAGCCCCGGTGTACAAGAAA 25
191	CGGTGCGACTTCTTACCTTCCTTTTAAACGAACCTGGACGTCGACGTCGACGACCTCGACGTCGACGTCGACGTCGACGTCGACGTCGACGTCGACGTCGACGTCGACGTCGACACCTCGACACACAC
	ATLKNGRKICLDLQAPLYKK -
	ATAATTAAGAAACTTTTGGAGAGTTAG (SEQ ID NO:34)
251	TATTAATTCTTTGAAAACCTCTCAATG
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Fig. 3C (SEQ ID NO:57)

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Fig. 3D (SEQ ID NO:59)

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21	L gagggtcgaggtcgtgtagtggtagtggga	180
	(SEQ ID NO:61) LQCLCVKTTSQVQPQHITSL	(58)
181	ggaggtgatcaaggccggaccccactgcccactgcccaactgatagccacgctgaagaa	240
	EVIKAGPHCPTAQLIATLKN	(78)
	tggacagaaaatttgcttggacctgcaagcccgctgtacpagaaaataattaagaaact	300
241	acctgtcttttaaacgaacctggacgttcggggfgacatgttcttttattaattctttga	
	G Q K I C L D L Q A P L Y K K I I K K L	(98)
301	tttggagagttag (SEQ ID NO:59)	313
	L E S (SEQ ID NO:68)	(101

Fig. 4A (SEQ ID NO:36)

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Fig. 4B (SEQ ID NO:41)

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Fig. 4C (SEQ ID NO:52)

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	aaat	tti	atca	aag	gaa	ctg	aga	gte	gati	gag	jagi	tgg:	acca	cac	tgo	gcc	aac	aca	gaa	att	12	9		
0	+	338	tagt	tt	ctt	gac	tet	ca	ct a	acto	CEC	acc	t gg t	gte	jacc	cgç	ttg	ıtgt	ctt	Laa				
	к	F	ı	ĸ	E	L	R	v	1	E	s	G	P	н	С	Α	N	т	E	1		•		
	att	qta	aag	cta	ago	gat	gg	aag	aga	gct	gtg	tct	gga	ecc	caa	gga	aaa	erg	ggr	geag	- 18			
30		car	rtc	gat	tc	ct	acc	ttc	tct	cga	cac	aga	cct	999	gtt	cct	ttt	gac	cca	cgc	-			
	ī	v	ĸ	L	s	D	G	R	E	L		: 1	. 0	P	K	Ε	N	W	٧	Q		-		
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190	+ -1							٠		ccc		rct I	aac	ıt.										
	l R	- 		 E		· ·	7 1		ĸ	R	A	E :	N :	s	(:	SEQ	ID	NO:	63)					

Fig. 4D (SEQ ID NO:54)

	IL-8M9-1
	IL-8M9-1 [36] [36] [36] [36] [36] [36] [36] [36]
1	gttcgaagtatactcacgatttctagacgcaacagtcacgtatttctgtatgaggtttggaaaggtgggg
	M S A K D L R C Q C I K T Y S K P F H P -
	ammatttatcamaagaactgagagtgastgagaggagcacacactgcgccamcacagamamatt 150
71	tttaaatagtttcttgactctcaccaaccottoo
	K F I K E L R V I E S G P H C A N T E I -
	attgtaaagctaagcgatggaagagagtgtgtctggaccccaaggaaaactgggtgcag 210
151	taacatttcgattcgctaccttctctcgatacaggar-19355
	IVKLSDGRELCLDPKERKVI
	agggttgtggagaagtttttgaagagggctgagaattca (SEQ ID NO:54)
211	agggtGtgggamau
	R V V E K F L K R A E N S - (SEQ ID NO:64)

11/25

Fig. 4E (SEQ ID NO:50)

						IL-	8М:	10-	1				1'	SEC	II C	nc	:51 ata	.) ctc	caa	acci	tt	cac	ccc	
	caago	tto	at	ato	gagt	gct	aa	aga	CCL	gaa	ctg 		greg										+	70
1	gttcg	aag	gti	CL	cacc	dict	LL	-95	,		-		-											
				м	s	A	H	ς :	D 1	LN	1 C	Q	С	1	K	Т	Y	s	K	₽	F	н	,	-
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71	ttta	aat	ag	ttt	CEE	gac	t c	tca	cta	acto	ctc	acc	t gg	tgt	gac	gcg	gtt	gtg	teti	ttaa				
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131	taac	ati	t t	ga:	tte	jcta	cc	tte	tct	cga	cac	aga	cct	ggs	get	cct	ttt	gac	cca	cát				
	1 '	, ,	к	L	s	D	G	R	Ε	L	c	L	D	P	ĸ	E	N	W	V	Ç	-			
								~=	agas	aaac	tq.	agaa	ttt	a	(SE	11	о и	0:50))					
191				-+-							***	rct I	aac	σt										
	R	v	v	E	к	F	L	ĸ	R	A	E	N	s		- (SEQ	ID	NO	:65)				

Fig. 4F

	· · · · · · · · · · · · · · · · · · ·	129
80	ATGAGTGCTAAAGAACTTAGATGTCAGTGCATAAAGACATACTCCAAACC	244
93	ATGAGTGCTAAAGAACTTAGATGTCAGTGCATTAGATGTCAGTGCATTAGATGTCAGTGCATTAGATGTCAGTGCATTAGATGTCAGTGCATTAGATGTCAGTGCATTAGATGTCAGTGCATTAGATGTCAGTGCAGTGCATTAGATGTCAGTGCAGTGCAGTGCATTAGATGTCAGTGCAGGAGTGCAGGAGAGGCAGAGTGCAGGAGAGAGGAGCAGAGAGAG	
20	TTTCCACCCCAAATTTATCAAAGAATACAGACGTATTGAGAGTGGACCAC	179
.50	TTTCCACCCCAAATTTATCAAAGAATACAGACGIIIGAGAGAGACGACGATTTATCAAAGAATACAGACGATATTGAGAGTGGACCACTTTCCACCCCAAATTTATCAAAGAATACAGACGTATTGAGAGTGGACCACTTTCCACCCCAAATTTATCAAAGAATACAGACGTATTGAGAGTGGACCAC	194
243	TTTCCACCCCAAATTATCAAAGA	
	ACTGCGCCAACACAGAAATTATTGTAAAGCTAAGCGATGGAAGAGAGCTG	229
100	ACTGCGCCAACAGAAATTATTGTAAAGCTAAGCGAGGAGAGAGGAGGAGGAGGAGGAAGAGAAATTATTGTAAAGCTAAGCGATGGAAGAAGAGAGAG	144
193	ACTGCGCCAACACAGAAATTATTGTAAAGCTAAGCGATGG	
	TGTCTGGACCCCAAGGAAAACTGGGTGCAGAGGGTTGTGGAGAAGTTTTT	279
230	TGTCTGGACCCCAAGGAAAACTGGTGCAGAGGGTTGTGGAGAAAGTTTTT TGTCTGGACCCCAAGGAAAACTGGGTGCAGAGGGGTTGTGGAGAAAGTTTTT	94
143		`
200	GAAGAGGGCTGAGAATTCATAGTAAGGATCC 310 (SEQID NO: SE)	,
200	GAAGAGGGCTGAGAATTCATTTTTTTTTTTTTTTTTTTT	
93	GAAGAGGCTGAGAATTCATAGTAAGGATCC 63	

acid sequence SAKDLRCQCIKTYSKPF-HPKFIKELRVIESGPHCANTEIIVKLSDGRELCLDPKENWVQRVVEKFLKRAENS (SEQ ID NO:48).

The invention additionally features a protein 5 having the amino acid sequence of a wild-type chemokine, e.g., IL-8, having four cysteine residues, with the following modifications: a) amino acids DLN as the 3 contiguous amino acids on the N-terminus side of and proximal to the first cysteine residue of the wild-type 10 chemokine; and b) amino acids $ELXVX_1X_2X_3X_4X_5X_6$ as the 10 amino acids on the N-terminus side of and proximal to the third cysteine of the wild-type chemokine, wherein ${\tt X}$ is any amino acid, e.g., R, N, E, or Q, and $X_1X_2X_3X_4X_5X_6$ can be any six consecutive amino acids on the N-terminus side 15 of and proximal to the third cysteine of the wild-type chemokine. In particular, the protein can have the amino acid sequence SAKDLNCQCIKTYSKPF-HPKFIKELRVIESGPHCANTEIIVKLSDGRELCLDPKENWVQRVVEKFLKRAENS

(SEQ ID NO:49).

The invention further features a protein having the amino acid sequence of a wild-type chemokine, e.g., 20 PF-4, having four cysteine residues, with the following modifications: a) amino acids ELR as the 3 contiguous amino acids on the N-terminus side of and proximal to the 25 first cysteine residue of the wild-type chemokine; and b) amino acids XDLQ as the 4 amino acids on the C-terminus side of and proximal to the fourth cysteine of the wildtype chemokine; wherein X can be any amino acid, e.g., L. In particular, the protein can have the amino acid

 ${\tt SAKELRCQCVKTTSQVRPRHITSLEVIKAGPHCPTAQLIATLKNGRKICLDLQAPLY}$ 30 sequence KKIIKKLLES (SEQ ID NO:22).

In addition, the invention features a protein having the amino acid sequence of a wild-type chemokine, 35 e.g., PF4, having four cysteine residues, with the

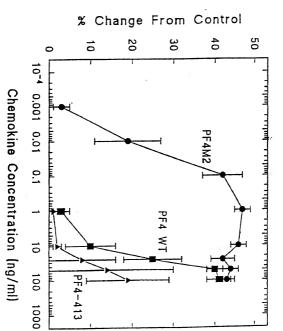


Fig. 6A

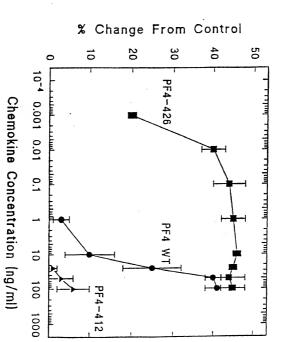
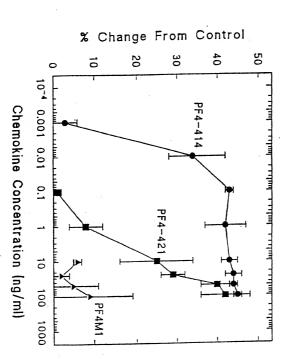


Fig. 6B

Fig. 6C





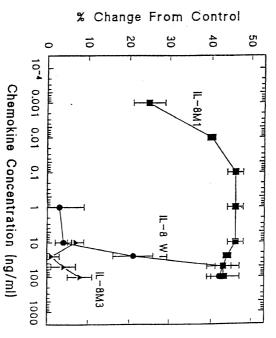
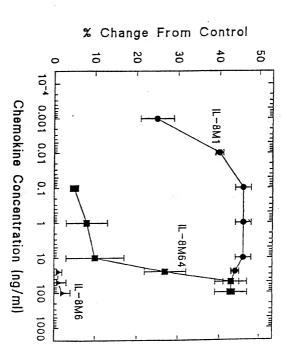


Fig. 6D

Fig. 6E



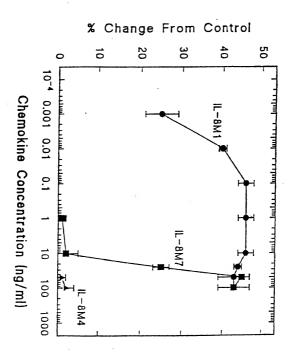


Fig. 6F

Fig. 6G

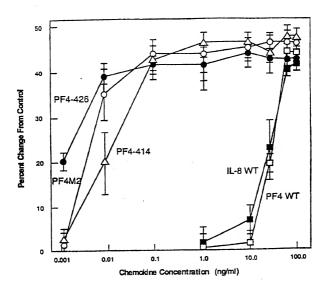


Fig. 6H

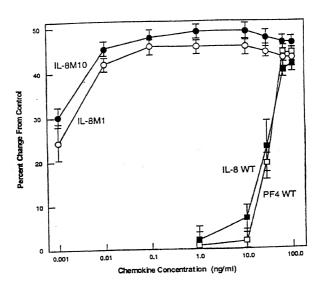


Fig. 7A

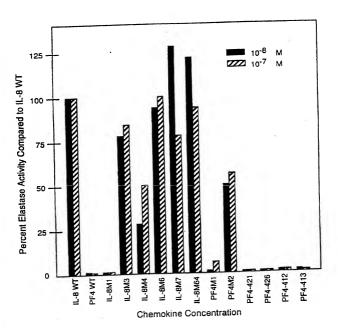


Fig. 7B

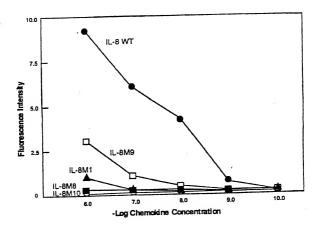
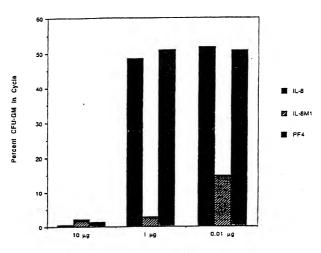
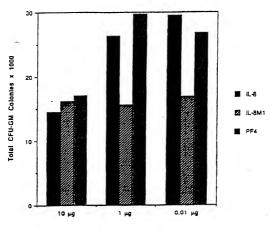


Fig. 8A



Chemokine Dose per Mouse

Fig. 8B



Chemokine Dose per Mouse

International application No. PCT/US95/13897

والشاء أووناهم والمعسر

CLAS	SIFICATION OF SUBJECT MATTER			
	112N 15/19 CO7K 14/52; Abik 30/19	IPC		
SCL:5	36/23.1; 530/300; 424/85.1; 930/140; 514/2 International Patent Classification (IPC) or to both nation	al classification and is C		
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	on searched other than minimum documentation to the exter	nt that such documents are included i	n the news searches	
	ata base consulted during the international search (name o	f data base and, where practicable,	search terms used)	
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Please Se	e Extra Sheet.			
	TO BE BELEVANT			
DOC	UMENTS CONSIDERED TO BE RELEVANT	rists of the relevant passages	Relevant to claim No.	
ategory*	Citation of document, with indication, where approp	, and, e	10-17, 19, 30-	
Y	PROCEEDINGS OF THE NATIONAL AC	CADEMY OF SCIENCES	34	
Y	PROCEEDINGS OF THE NATIONAL ACTION OF THE NAT	C-LEWIS ET AL Plateics		
	Factor 4 Binds to Interlegality is	Modified with Glu-Leu-		
	Neutrophils when its N Terminus is Arg", pages 3574-3577, see page	3574, column 1, see		
	Arg", pages 35/45377, 555 F. abstract.			
		. A. J. 10. June 1993, see	1-9, 17-18, 20-	
Α	WO, A, 93/11159 (CLARK-LEWIS ET AL) 10 June 1993, see 1-9, 17-18, 20 24, 27-34			
	entire document.			
_	THE JOURNAL OF IMMUNOLOGY,	Volume 149, Number 3,	1-24, 27-34	
Α	THE JOURNAL OF IMMUNOLOGY, Issued 01 August 1992, MAZE ET	AL "Myelosuppressive		
	issued 01 August 1992, MAZE ET Effects in vivo of Purified Recombin	1009 see page 1008,		
	Inflammatory Protein- 1a , pages 100	14-1000, 000 F-0		
	column 1, Figure 6.			
		See patent family annex.		
X Fu	rther documents are listed in the continuation of Box C.		international filing date or priority	
•	of sized documents:	date and not in contract underlying the	invention	
٠٨٠	Special categories of class considered document defining the general state of the art which is not considered to be part of particular relevance	X* document of particular relevance	; the claimed invention cannot be aidered to involve an inventive step	
·E•	earlier document published on or after the international trials in	when the document is taken are	and the second be	
.r.	cited to establish the publication	COMPRESSED IN THE CASE OF THE	s; the claimed invention cannot be stive step when the document is such documents, such combination in the art	
٠٥٠	special reason (as specified) document referring to an oral disclosure, use, exhibition or other	being obvious to a person skilled	in the art	
1	means document published prior to the international filing date but later than	·& document member of the same p	atent family	
·P	the priority date claimed the actual completion of the international search	Date of maining of the another	search report	
1		29 FEB 1996		
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Comm	(ISSICIALI OF FEIGURE	Telephone No. (703) 308-0196	<u> </u>	
Washington, D.C. avan		1 telephone ivo.	141	

International application No.
PCT/US95/13897

(Continual	ion). DOCUMENTS CONSIDERED TO BE RELEVANT	Relevant to claim No.
ategory*	Citation of document, with indication, where appropriate, of the relevant passages	
A	EXPERIMENTAL HEMATOLOGY, Volume 22, issued 1994, COOPER ET AL "Myelosuppressive Effects in Vivo With Very Low Dosages of Monomeric Recombinant Murine Macrophage Inflammatory Protein-la", pages 186-193, see page 187, Figure 1.	1-24, 27-34
A	GENE, Volume 151, issued 1994, JOHNSON II et al "Cloning of Two Rabbit GRO Homologues and Their Expression in Alveolar Macrophages", pages 337-338, see page 338, see Figure 1.	1-19, 27-34
A	GENE, Volume 151, issued 1994, POWER ET AL "Cloning of a Full-length cDNA Encoding the Neutrophil-activating Peptide FNA-78 from Human Platelets", pages 333-334, see page 334, see Figure 1.	1-19, 27-34
	-	*
		- A

Form PCT/ISA/210 (continuation of second sheet)(July 1992)*

International application No. PCT/US95/13897

والشروا ويتأثيه والمعاصرة

Box 1 Observations where certain claims were found unsearchable (Continuation	n of item 1 of first sheet)					
Box 1 Observations where certain Campaigness of certain claims under Article 17(2)(a) for the following reasons: This international report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:						
Claims Nos.: because they relate to subject matter not required to be scarched by this Au	sthority, namely:					
 Claims Nos.: because they relate to parts of the international application that do not comply an extent that no meaningful international search can be carried out, specified. 	y with the prescribed requirements to such fically:					
Claims Nos.: because they are dependent claims and are not drafted in accordance with the	second and third sentences of Rule 6.4(a).					
Box II Observations where unity of invention is lacking (Continuation of item	2 of first sheet)					
Box II Observations where using our multiple inventions in this international This International Searching Authority found multiple inventions in this international	l application, as follows:					
Please See Extra Sheet.	1					
As all required additional search fees were timely paid by the applicant, thi	is international search report covers all scarchable					
claims.	disional fee, this Authority did not invite payment					
claims. 2. As all scarchable claims could be searched without effort justifying an ad-	ditional rec. discrete					
of any additional Ice.						
As only some of the required additional scarcifical scale only those claims for which fees were paid, specifically claims Nos.:						
	*					
No required additional search fees were timely paid by the applicant. restricted to the invention first mentioned in the claims; it is covered to 1-24 and 27-34.	Consequently, this international search report is y claims Nos.:					
	d by the applicant's protest.					
Remark on Protest The additional search fees were accompanied	itional search foca.					
Remark on Protest No protest accompanied the payment of add	Mount seems					

Form PCT/ISA/210 (continuation of first sheet(1))(July 1992)*

International application No. PCT/US95/13897

entropy of the and the

Electronic data bases consulted (Name of data base and where practicable terms used):

APS, CAS ONLINE, MEDLINE, BIOSIS, SCISEARCH search terms: chemokines, interleukin 8, platelet factor 4, mutants, modifications, derivatives, analogs, treatment, therapy, administration.

BOX II. OBSERVATIONS WHERE UNITY OF INVENTION WAS LACKING

This ISA found multiple inventions as follows:

This application contains the following inventions or groups of inventions which are not so linked as to form a single inventive concept under PCT Rule 13.1. In order for all inventions to be examined, the appropriate additional

1. Claims 1-24 and 27-34, drawn to a modified chemokine, a nucleic acid encoding the modified chemokine, and the use of the modified chemokine for the manufacture of a medicament for suppressing proliferation of an actively dividing

II. Claims 25-26, drawn to a method of detecting and isolating CD34+ myeloid cells with a modified chemokine.

The inventions listed as Groups I-II do not relate to a single inventive concept under PCT Rule 13.1 because, under PCT Rule 13.2, they lack the same or corresponding special technical features for the following reasons:

The processes of Groups I-II are separate and distinct inventions in that the use of the modified chemokine for the manufacture of a medicament for suppressing proliferation of an actively dividing myeloid cell of Group I and a method of detecting and isolating CD34+ mycloid cells of Group II are practiced with materially different process steps which constitute the special technical (catures which define the contribution of each invention. Since these special technical features are not shared by each invention, the inventions of Groups I-II do not form a single inventive concept within the meaning of Rule 13.2.

Form PCT/ISA/210 (extra sheet)(July 1992)*